



PATHOGEN DIAGNOSIS BY METAGENOMIC NANOPORE SEQUENCING: A REVIEW

Nataraja B.T.¹, Rathna Kumari B.M.², Mohan Kumar B.S.³ and Uddappanda Bopaiah Roy^{4*}

¹Department of Microbiology, Government Science College, Chitradurga, Karnataka, India

²Department of Botany, Government First Grade College, Vijayanagara, Bengaluru, Karnataka, India

³Department of Zoology, Maharani's Science College for Women, Bengaluru, Karnataka, India

⁴Department of Zoology, Government Science College, Bengaluru, Karnataka, India

***Corresponding Author: Dr. Uddappanda Bopaiah Roy**

Email: royub09@gmail.com

Abstract

All microorganisms contain genetic material (DNA or RNA), thereby making sequencing a useful technique for the identification of microbes. Identification of microorganisms in its natural environment is known as metagenomics. The expense of next generation sequencing has been reduced by a certain size since its invention in 2004, but soon developed as a powerful molecular technique for identification and characterization of microbes from clinical samples from patients. Nanopore Sequencing, a next generation sequencing approach has proved as an innovative molecular technique that upgrades the capacity to identify pathogens. The technique involves the sequencing of a single strand using protein nanopores and adaptable electronic gadgets. The current review deals with how nanopore sequencing plays an important role in metagenomics and few real time examples on how it enables to reduce the consumption of antibiotics.

Keywords: Metagenomics, Nanopore Sequencing, Next Generation Sequencing, Pathogens, Antibiotics.

INTRODUCTION

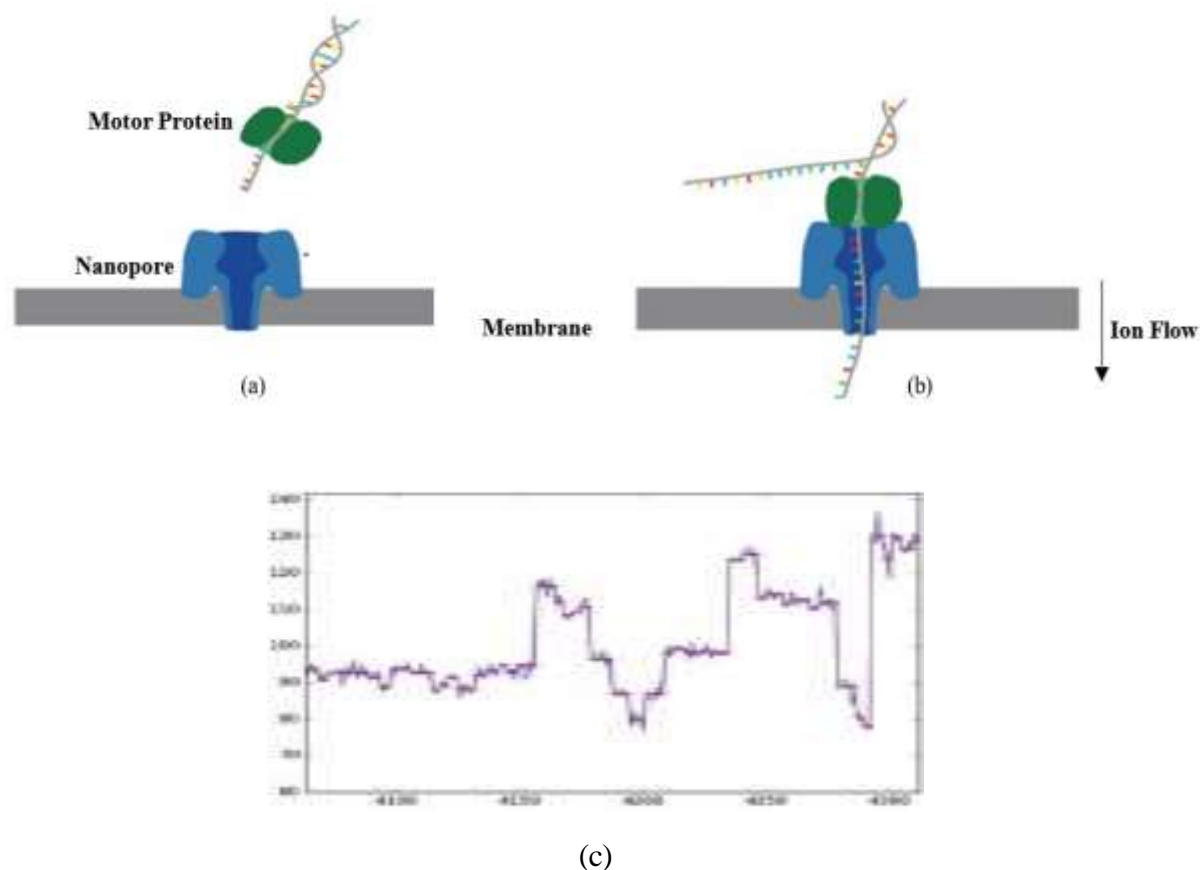
Study of genomes recuperated from natural samples instead of those from clonal societies is known as Metagenomics. Metagenomics is also known as Environmental Genomics or Community Genomics [1]. The term is derived from the factual idea of “meta” examination (the procedure of measurably joining separate investigation) and genomics (the study of an organism's hereditary material basically in the context of uncultured microorganisms).

This moderately new field of hereditary research permits the genomic investigation of life forms that are not effortlessly cultured in lab. The aggregate genomes of organisms indigenous to specific natural surroundings particularly of outrageous atmospheres, for example, hot springs, salt lakes or high heights are presently regularly alluded to as the metagenome [2]. Metagenomics is utilized as a method for deliberately examining, ordering and controlling the whole hereditary material segregated from a specific ecological example. Research on metagenome starts with the extraction of DNA from natural examples and experiences the investigation of libraries arranged by cloning in suitable vectors. This methodology expands on late advances in microbial genomics, PCR enhancement and cloning DNA that has been extricated legitimately from the soil or some other ecological examples and

cloning of quality that offer sequence similarity [3].

In 1977 Fred Sanger et al. published two methodology-based papers with respect to the fast assurance of DNA sequences that assisted with changing science and gave another apparatus to decoding total qualities and later the whole genome [3-5]. The strategies drastically improved existing DNA sequencing procedures created by Maxam and Gilbert distributed around the same time and Sanger and Coulson's own "add and less" strategy distributed 2 years sooner [6,7]. The favorable position of managing less harmful synthetic compounds and radioisotopes made "Sanger sequencing" the main DNA sequencing technique utilized for the following 30 years. The gel-based sequencing innovation has experienced sensational improvement in throughput from one parallelization, mechanization, and refinement of sequencing strategies and science. Ongoing advances in microfibration have brought about enhancements of Sanger sequencing by multiplexing and scaling down [8]. In spite of numerous upgrades, the gel-based Sanger sequencing innovation despite everything faces downsides for the sake of cost and low throughput. For accomplishing high throughput, numerous business organizations and logical labs have thought of various methods for high throughput sequencing with a sensible expense. The following technologies are known as next generation sequencing techniques: Sequence by synthesis, Sequencing by ligation; Sequencing by hybridization; Single molecule DNA sequencing; Nanopore sequencing and Multiplex polony sequencing. Next generation sequencing has effects on genetic applications like metagenomics, comparative genomics, high throughput polymorphism identification, examination of small RNAs, mutation screening, transcriptome profiling, methylation profiling and chromatin remodeling. The significant estimations for the accomplishment of the next generation technology are sequence (read length), sequence quality, high throughput, and low cost. Nanopore sequencing was invented by David Deamer at the University of California Santa Cruz, and by George Church and Daniel Branton (both at Harvard University). Starting in the mid-1990s, scholastic research centers arrived at a progression of achievements towards building up a practical nanopore sequencing stage [9,10]. These achievements included the translocation of individual nucleic acid strands in single record order [11], processive enzymatic control of DNA at single-nucleotide precision [12] and the accomplishment of single-nucleotide resolution [13,14]. Single nucleotide polymorphism can be recognized by a change in the limit voltage of a nanopore [15].

In 2014, the MinION, the commercial sequencer utilizing nanopore innovation, was released by Oxford Nanopore Technologies (ONT). MinION recognizes DNA bases by estimating the progressions in electrical conductivity created as DNA strands go through an organic pore. Its movability, moderateness, and speed in information creation makes it reasonable for continuous applications, the arrival of the since quite a while ago read sequencer MinION has in this manner produced a lot of fervor and enthusiasm for the genomics network. A biological nanopore is incorporated into an electrically resistant artificial membrane and a voltage is applied across the membrane. DNA molecules are set up as per one of a lot of standard library readiness conventions which include connecting a leader adaptor and motor protein to one strand of DNA. During sequencing, the motor protein unfastens dsDNA and passes a solitary strand through the pore a base at once. The nearness of the DNA particle in the pore causes a redirection in the current over the pore and this present change can be identified with the specific bases present in the pore at a place that is normally dependent on 5-mer or 6-mer models. During a test, each nanopore in the array analyses molecules in the sample autonomously of the different nanopores. The most limited time to begin gathering exploratory information is the time taken for one analyte particle to effectively associate with one nanopore in the cluster. While a solitary DNA atom may take milliseconds to seconds to go through the nanopore, information begins opening up when it begins going through the pore. At a place where a DNA particle has gone through a nanopore, another will load. Data investigation takes place continuously as information streams from the sensor chip in equal from numerous nanopores. Accordingly, a more drawn-out run empowers more information focuses to be gathered, more certainty about a perception to be accomplished, greater estimation precision to be gotten, and a more prominent scope of examinations to happen [16].



- (a) A biological nanopore is inserted into an electrically resistant synthetic membrane. A potential is applied across the membrane, resulting in ion flow. Library DNA molecules have adaptors with aliphatic tethers which preferentially locate to the membrane for a localized library concentration.
- (b) The motor protein bound to the other adaptor docks with the pore, and passes the DNA molecule through it.
- (c) Bases in the nanopore cause disruptions in the current which are characteristic of their sequence.

ADVANTAGES OF mNGS IN PATHOGEN DETECTION

The etiology of suspected infections in intensely poorly hospitalized patients regularly stays undiscovered, bringing about postponed or insufficient treatment, delayed stays, readmissions, and expanded mortality and bleakness [17, 18]. As often, these patients are immunocompromised because of disease, genetic disorders, or transplantation, particularly on the off chance that they are in tertiary consideration clinical focuses, making them incredibly powerless against contaminations. In this setting, the causative specialist can incorporate various both normal and unprecedented pathogens, going from viruses to bacteria, fungi and parasites. Hypothesis driven molecular testing, for example, PCR can include various individual tests for explicitly focused on targeted organisms, but may miss an uncommon pathogen or use groundworks containing bungles to the microbial strain included, which diminishes the affectability of location [19]. A hypothesis free symptomatic methodology that can possibly distinguish almost any living being would prompt a sensational change in outlook in microbial indicative testing.

In contrast with other analytic innovations, mNGS offers various points of interest: A central bit of leeway of mNGS is fair examining, which empowers wide recognizable proof of referred to just as unexpected pathogens or even the revelation of new life forms [20]. mNGS can likewise be coupled to focused methodologies, for example, the utilization of groundworks from saved 16S ribosomal RNA (rRNA) and interior interpreted spacer groupings for, individually, general bacterial and

contagious location [21, 22], which can take into consideration species level distinguishing proof of these life forms. Another favorable position of mNGS is that it can give the helper genomic data vital for developmental following [23], strain distinguishing proof [24,25] and expectation of medication obstruction [26]. NGS can give quantitative or semiquantitative information with respect to the grouping of living beings in the example through the checking of sequenced peruses, which is valuable for polymicrobial tests or in cases in which more than one pathogen has been ensnared in the ailment procedure [22].

THE CONCEPT OF NANOPORE SEQUENCING

The possibility of nanopore sequencing was proposed by Deamer and Branton and freely by Church [27]. The idea is that if bases could actuate distinctive ionic current blasts during DNA navigating through a little channel, at that instance it would turn into an absolutely new sequencing strategy. In 1993, Deamer, Branton, and Kasiannowicz utilized α -hemolysin (α HL), a poisonous pore shaping protein discharged by *Staphylococcus aureus* to assault a lipid bilayer, to identify DNA translocation through α -HL nanopore [28]. In 1996, their aftereffects of DNA translocation through α -HL nanopore was published [29].

Bayley and associates announced that α -HL is a 232.4 kDa layer channel protein [30]. Their crystal structure examination of α -HL uncovered a \sim 10 nm-high empty mushroom- formed homoheptamer complex containing a \sim 10 nm-wide extra membranal top and a \sim 5.2 nm-long transmembrane β -barrel stem [31]. The base distance across at the narrowing site of the channel is \sim 1.4 nm, which is associated with the β -barrel with the vestibule of 2.6 nm in measurement at the trans side.

A α -HL nanopore is embedded into a lipid bilayer which isolates little volumed chambers, each associated with a cathode and an anode of a fix cinch speaker (PCA). The \sim 1.4 nm tightening of α -HL pore permits just individual single-abandoned DNA (ssDNA) or RNA other than 2 nm-thick double-stranded DNA (dsDNA) to cross through. Various bases along the adversely charged DNA strand will cause electric flow changes over the span of translocating through the nanopore under an applied electric field. On the off chance that the variances are base-explicit, these electric signs or marks can be in the long run changed over into DNA sequence information [32].

Viruses is an intricate ailment phenotype where the basic microbiome could impact grimness and mortality. Amplicon and metagenomic MinION based sequencing was utilized to quickly (inside 8 hours) recognize any virus and evaluate the microbiome in nasopharyngeal swabs of the patients with target virus. The research discovered that utilizing this methodology coupled to MinION based sequencing uncovered the hereditary information within ten hours.

Illumina sequencing is viewed as the best quality level for the research of microbiomes, but it is restricted by getting just short DNA arrangements to dissect. As another option, Oxford Nanopore Technologies (ONT) had built up another sequencing procedure dependent on nanopores that can be completed in the MinION, a versatile gadget with a low introductory cost which long DNA readings can be acquired with. The purpose of the investigation was to analyze the presentation of the two kinds of sequencing applied to tests of ruminal content utilizing a comparable pipeline. The ONT sequencing gave comparative outcomes to the Illumina sequencing, in spite of the fact that it had the option to order a greater number of readings at the species level, potentially because of the expansion in the read size. The outcomes additionally propose that, because of the size of the peruses, it is conceivable to get a similar measure of data in fewer hours. Be that as it may, identification of archaeal and eukaryotic species is still hard to achieve because of their low bounty in the rumen contrasted with microorganisms, proposing various pipelines and systems are expected to get an entire portrayal of the less plenteous species in the rumen microbiota [33].

The ability of the MinION sequencing way to deal with the identification and describing infections contaminating a water sweet potato plant was surveyed. The sequencing stage reliably uncovered the nearness of a few plant infection animal types, including *Dioscorea bacilliform* infection, Yam gentle mosaic infection and Yam chlorotic putrefaction infection. A conceivably novel ampelovirus was additionally distinguished by a complimentary Illumina sequencing approach. The full-length genome grouping of sweet potato chlorotic putrefaction infection was resolved utilizing Sanger sequencing, which empowered assurance of the inclusion and sequencing exactness of the MinION innovation. While the complete mean sequencing blunder pace of sweet potato chlorotic rot infection related MinION peruses was 11.25%, it was demonstrated that the agreement succession got either by anew get together or in the wake of mapping the MinION peruses on the infection genomic arrangement was >99.8% indistinguishable with the Sanger- inferred reference grouping. From the viewpoint of potential plant illness symptomatic utilizations of MinION sequencing, these degrees of sequencing exactness show that the MinION approach can be utilized to both dependably distinguish and precisely succession almost full-length positive-sense single-strand polyadenylated RNA plant infection genomes [34].

With its little size and minimal effort, the hand-held MinION sequencer is a useful asset for in-field observation. Utilizing a metagenomic approach, it permits non-focused on identification of infections in an example inside a couple of hours. The investigation intended to decide the capacity of the MinION to metagenomically identify and portray an infection from a tainted mosquito. RNA was extricated from an *Aedes notoscriptus* mosquito contaminated with Ross River infection (RRV), changed over into cDNA and sequenced on the MinION. Bioinformatic examination of the MinION peruses prompted location of full-length RRV, with peruses of up to 2.5 kb adding to the get together. The cDNA was additionally sequenced on the MiSeq sequencer, and the two stages recuperated the RRV genome with >98% exactness. This evidence of idea study showed the metagenomic discovery of an arbovirus, utilizing the MinION, straightforwardly from a mosquito with insignificant sample purification [35].

The acquaintance of metagenomic sequencing with diagnostic microbiology has been hampered by gradualness, cost and multifaceted nature. It was investigated whether MinION nanopore sequencing could quicken finding and obstruction profiling, utilizing entangled urinary tract diseases as a model. Bacterial DNA was improved from clinical pees (n=10) and from sound pees 'spiked' with multiresistant *Escherichia coli* (n=5), and then sequenced by MinION. Sequences were investigated utilizing outer databases and bioinformatic pipelines or, at last, utilizing coordinated continuous examination applications. Results were contrasted and Illumina information and opposition phenotypes. MinION accurately recognized pathogens without culture and, from 55 procured obstruction qualities identified in the developed microscopic organisms by Illumina sequencing, 51 were found by MinION sequencing straightforwardly from the urines; with three of the four disappointments in an early run with low genome inclusion. Opposition giving changes and allelic variations were not dependably distinguished. MinION sequencing thoroughly distinguished pathogens and obtained opposition qualities from pee in a time span of 4hrs from test to result [36].

Foodborne episodes of *Salmonella* stay a squeezing general wellbeing concern. An enormous flare-up of *Salmonella* enterica serovar Enteritidis phage type 14b influencing in excess of 30 patients in an emergency clinic was distinguished. This flare-up was connected to network, national and European-wide cases. Emergency clinic patients with *Salmonella* were at high hazard, and required a quick reaction. First this episode was examined by entire genome sequencing utilizing a novel quick convention on the Illumina MiSeq; and afterward, coordinated this information with entire genome information from observation sequencing and setting the flare-up in a national setting. Additionally, the capability of a recently discharged sequencing innovation, the MinION from Oxford Nanopore Technologies in the administration of a medical clinic flare-up of *Salmonella* was investigated. It was shown that that quick MiSeq sequencing can diminish an opportunity to answer contrasted with the

standard sequencing convention with no effect on the outcomes. It was shown that the MinION can secure clinically applicable data continuously and within minutes of a DNA library being loaded [37].

Metagenomics is a significant subject for the investigation of microbial networks, but has been constrained by the trouble of "binning" the subsequent successions into bunches relating to the individual species and strains that establish the network. Additionally, there are directly no techniques to follow the progression of portable DNA components, for example, plasmids through networks or to figure out which of these are co-restricted inside a similar cell. We address these constraints by applying Hi-C, an innovation initially intended for the investigation of three dimensional genome structure in eukaryotes, to gauge the cell co confinement of DNA successions. It was utilized that the Hi-C information created from a basic engineered metagenome test to precisely bunch metagenome gathering contigs into bunches that contain almost complete genomes of every specie. The Hi-C information likewise dependably connected plasmids with the chromosomes of their host and with one another. It was additionally shown that Hi-C information gives a long-run sign of strain-explicit genotypes, demonstrating such information might be valuable for high-goals genotyping of microbial populaces. The work showed that Hi-C sequencing information give important data to metagenome investigations that are not as of now realistic by different techniques. This metagenomic Hi-C strategy could encourage future investigations of the fine-scale populace structure of organisms, just as investigations of how anti-toxin obstruction plasmids (or other hereditary components) activate in microbial networks. The strategy isn't restricted to microbiology; the hereditary engineering of different heterogeneous populaces of cells could likewise be concentrated with this method [38].

Next generation sequencing has extraordinarily extended our capacity to inquiry the identity and hereditary piece of whole networks of microbial life forms. This zone of research, known as metagenomics, doesn't depend after refined the individual life forms. Or maybe, the hereditary material from the whole network is handled and sequenced real time. From this grouping information, scientists can decide the overall populace of living beings inside the network just as figure out which qualities and metabolic pathways are available and communicated in the microbial network. While these procedures have been applied to a wide scope of ecological examples, metagenomics is additionally the focal point of escalated look into on human-related microbial networks. The extent of these human metagenomics examines are very changed, but still all have a shared objective of endeavoring to comprehend the significant job that human commensal microbial networks play in wellbeing and malady. The early outcomes from considering the human metagenome demonstrate a fundamental job that microbial networks play in resistance, wellbeing, and ailment [39].

High-throughput next generation sequencing (NGS) has been rapidly adjusted into numerous parts of biomedical research and started to connect with the clinical practice. The latter viewpoint will empower the use of genomic information into clinical practice in this and one decade from now and will significantly change the determination, visualization and treatment of numerous human ailments. It will additionally request both philosophical and clinical educational program changes in the preparation of our future doctors. Be that as it may, critical groups should be defeated before an extreme utilization of NGS in genomic medication can be functional and productive.

New maladies keep on developing in both human and animal populaces, and the significance of creatures, as supplies for infections that can cause zoonoses are obvious. In this manner, an increased information on the viral flora in animals, both in healthy and ailing people, is significant both for animal and human wellbeing. Viral metagenomics is a culture-free methodology that is utilized to explore the total viral hereditary populaces of an example. There are various potential strides of a viral metagenomic study using grouping free intensification, high-throughput sequencing, and bioinformatics to recognize infections. With this innovation, various infections can be identified at the same time and novel and profoundly unique infections can be found and hereditarily described just because. The utilization of viral metagenomics in veterinary science has considerably expanded

during the previous decade through the progression of new sub-atomic instruments and through an expanded comprehension of the significance of animal viruses to both animal and human welfare [40]

CONCLUSION

Nanopore sequencing innovations have been advancing at an extraordinarily quick speed while the second gen sequencers have been making ready for sequencing-based customized medication. It very well may be imagined that novel sequencing ideas and advancements, including strategies/materials for nanopore manufacture, parallelization procedures for nanopore exhibits and identification, and speed control of DNA translocation, and so on., will encourage the improvement of legitimately perusing nanopore sequencers that by and by depend on aberrant perusing. It can likewise be anticipated that a definitive objective of clinical sequencing will be accomplished soon alongside the development in sequencing innovation and grouping annotation.

Several patients may require various sequencing to screen genome strength during the concerned treatment. Regardless of whether cost for each sequencing and translation is \$1000, it is as yet costly for standard families. Accordingly, how to promote sequencing- based customized medication will require joint endeavors from different fronts.

REFERENCES

1. Handelsman, J et al. "Molecular biological access to the chemistry of unknown soil microbes: a new frontier for natural products." *Chemistry & biology* vol. 5,10 (1998): R245-9. doi:10.1016/s1074-5521(98)90108-9
2. Rondon, M R et al. "Cloning the soil metagenome: a strategy for accessing the genetic and functional diversity of uncultured microorganisms." *Applied and environmental microbiology* vol. 66,6 (2000): 2541-7. doi:10.1128/AEM.66.6.2541-2547.2000
3. Sanger, F., Air, G., Barrell, B. *et al.* Nucleotide sequence of bacteriophage ϕ X174 DNA. *Nature* **265**, 687–695 (1977). <https://doi.org/10.1038/265687a0>
4. Liles, Mark R et al. "A census of rRNA genes and linked genomic sequences within a soil metagenomic library." *Applied and environmental microbiology* vol. 69,5 (2003): 2684-91. doi:10.1128/AEM.69.5.2684-2691.2003
5. Maxam, A M, and W Gilbert. "A new method for sequencing DNA." *Proceedings of the National Academy of Sciences of the United States of America* vol. 74,2 (1977): 560-4. doi:10.1073/pnas.74.2.560
6. Air, G M et al. "Nucleotide and amino acid sequences of gene G of omegaX174." *Journal of molecular biology* vol. 108,3 (1976): 519-33. doi:10.1016/s0022-2836(76)80134-9
7. Metzker, Michael L. "Emerging technologies in DNA sequencing." *Genome research* vol. 15,12 (2005): 1767-76. doi:10.1101/gr.3770505
8. Branton, D., Deamer, D., Marziali, A. *et al.* The potential and challenges of nanopore sequencing. *Nat Biotechnol* **26**, 1146–1153 (2008). <https://doi.org/10.1038/nbt.1495>
9. Deamer, D., Akeson, M. & Branton, D. Three decades of nanopore sequencing. *Nat Biotechnol* **34**, 518–524 (2016). <https://doi.org/10.1038/nbt.3423>
10. Kasianowicz JJ, Brandin E, Branton D, Deamer DW. 1996. Characterization of individual polynucleotide molecules using a membrane channel. *Proc Natl Acad Sci U.S.A.* 1996, 93(24): 13770-13773, <https://doi.org/10.1073/pnas.93.24.13770>
11. Cherf, G., Lieberman, K., Rashid, H. *et al.* Automated forward and reverse ratcheting of DNA in a nanopore at 5-Å precision. *Nat Biotechnol* **30**, 344–348 (2012). <https://doi.org/10.1038/nbt.2147>
12. Ayub, M., & Bayley, H. (2012). Individual RNA base recognition in immobilized oligonucleotides using a protein nanopore. *Nano letters*, 12(11), 5637–5643. <https://doi.org/10.1021/nl3027873>
13. Manrao, E., Derrington, I., Laszlo, A. *et al.* Reading DNA at single-nucleotide resolution with a mutant MspA nanopore and phi29 DNA polymerase. *Nat Biotechnol* **30**, 349–353 (2012). <https://doi.org/10.1038/nbt.2171>

14. Zhao, Q et al. "Detecting SNPs using a synthetic nanopore." *Nano letters* vol. 7,6 (2007): 1680-5. doi:10.1021/nl070668c
15. Hoenen, Thomas et al. "Nanopore Sequencing as a Rapidly Deployable Ebola Outbreak Tool." *Emerging infectious diseases* vol. 22,2 (2016): 331-4. doi:10.3201/eid2202.151796
16. Bleeker-Rovers, Chantal P et al. "A prospective multicenter study on fever of unknown origin: the yield of a structured diagnostic protocol." *Medicine* vol. 86,1 (2007): 26-38. doi:10.1097/MD.0b013e31802fe858
17. Ewig, S et al. "Factors associated with unknown aetiology in patients with community-acquired pneumonia." *The European respiratory journal* vol. 20,5 (2002): 1254-62. doi:10.1183/09031936.02.01942001
18. Wilson, Michael R et al. "Actionable diagnosis of neuroleptospirosis by next-generation sequencing." *The New England journal of medicine* vol. 370,25 (2014): 2408-17. doi:10.1056/NEJMoa1401268
19. Chiu, Charles Y. "Viral pathogen discovery." *Current opinion in microbiology* vol. 16,4 (2013): 468-78. doi:10.1016/j.mib.2013.05.001
20. Cummings, Lisa A et al. "Clinical Next Generation Sequencing Outperforms Standard Microbiological Culture for Characterizing Polymicrobial Samples." *Clinical chemistry* vol. 62,11 (2016): 1465-1473. doi:10.1373/clinchem.2016.258806
21. Salipante, Stephen J et al. "Coinfection of *Fusobacterium nucleatum* and *Actinomyces israelii* in mastoiditis diagnosed by next-generation DNA sequencing." *Journal of clinical microbiology* vol. 52,5 (2014): 1789-92. doi:10.1128/JCM.03133-13
22. Gire, Stephen K et al. "Genomic surveillance elucidates Ebola virus origin and transmission during the 2014 outbreak." *Science (New York, N.Y.)* vol. 345,6202 (2014): 1369-72. doi:10.1126/science.1259657
23. Salipante, Stephen J et al. "Application of whole-genome sequencing for bacterial strain typing in molecular epidemiology." *Journal of clinical microbiology* vol. 53,4 (2015): 1072-9. doi:10.1128/JCM.03385-14
24. Deurenberg, Ruud H et al. "Application of next generation sequencing in clinical microbiology and infection prevention." *Journal of biotechnology* vol. 243 (2017): 16-24. doi:10.1016/j.jbiotec.2016.12.022
25. Sahoo, M. K., Lefterova, M. I., Yamamoto, F., Waggoner, J. J., Chou, S., Holmes, S. P., Anderson, M. W., & Pinsky, B. A. (2013). Detection of cytomegalovirus drug resistance mutations by next-generation sequencing. *Journal of clinical microbiology*, 51(11), 3700–3710. <https://doi.org/10.1128/JCM.01605-13>
26. Pennisi, E. Search for pore-fection. *Science* 2012, 336(6081): 534–537. DOI: 10.1126/science.336.6081.534
27. Song, L et al. "Structure of staphylococcal alpha-hemolysin, a heptameric transmembrane pore." *Science (New York, N.Y.)* vol. 274,5294 (1996): 1859-66. doi:10.1126/science.274.5294.1859
28. Kasianowicz, J. J., Brandin, E., Branton, D., and Deamer, D. W. (1996). Characterization of individual polynucleotide molecules using a membrane channel. *Proc. Natl. Acad. Sci. U.S.A.* 93, 13770–13773.
29. Gouaux, J E et al. "Subunit stoichiometry of staphylococcal alpha-hemolysin in crystals and on membranes: a heptameric transmembrane pore." *Proceedings of the National Academy of Sciences of the United States of America* vol. 91,26 (1994): 12828-31. doi:10.1073/pnas.91.26.12828
30. Gouaux, E. "alpha-Hemolysin from *Staphylococcus aureus*: an archetype of beta-barrel, channel-forming toxins." *Journal of structural biology* vol. 121,2 (1998): 110-22. doi:10.1006/jsbi.1998.3959
31. Deamer, David. "Nanopore analysis of nucleic acids bound to exonucleases and polymerases." *Annual review of biophysics* vol. 39 (2010): 79-90. doi:10.1146/annurev.biophys.093008.131250

32. Bayley, Hagan. “Sequencing single molecules of DNA.” *Current opinion in chemical biology* vol. 10,6 (2006): 628-37. doi:10.1016/j.cbpa.2006.10.040
33. Eme, L., Spang, A., Lombard, J. *et al.* Archaea and the origin of eukaryotes. *Nat Rev Microbiol* **15**, 711–723 (2017). <https://doi.org/10.1038/nrmicro.2017.133>
34. Filloux, Denis *et al.* “Nanopore-based detection and characterization of yam viruses.” *Scientific reports* vol. 8,1 17879. 14 Dec. 2018, doi:10.1038/s41598-018-36042-7
35. Batavska J. *et al.*, “Metagenomic arbovirus detection using MinION nanopore sequencing”. *Journal of Virological Methods*. 2018, 249: 79-84. <https://doi.org/10.1016/j.jviromet.2017.08.019>
36. Jain, M., Olsen, H.E., Paten, B. *et al.* The Oxford Nanopore MinION: delivery of nanopore sequencing to the genomics community. *Genome Biol* **17**, 239 (2016). <https://doi.org/10.1186/s13059-016-1103-0>
37. Finn, S., Condell, O., McClure, P., Amézquita, A., & Fanning, S. (2013). Mechanisms of survival, responses and sources of Salmonella in low-moisture environments. *Frontiers in microbiology*, 4, 331. <https://doi.org/10.3389/fmicb.2013.00331>
38. Lawson, C.E., Harcombe, W.R., Hatzenpichler, R. *et al.* Common principles and best practices for engineering microbiomes. *Nat Rev Microbiol* **17**, 725–741 (2019). <https://doi.org/10.1038/s41579-019-0255-9>
39. Song, S., Jarvie, T., Hattori, M. Our. Second Genome—Human Metagenome: How Next-Generation Sequencer Changes our Life Through Microbiology- Chapter Three - Advances in Microbial Physiology, 2013, 62: 119-144, <https://doi.org/10.1016/B978-0-12-410515-7.00003-2>
40. Blomström, Anne-Lie. “Viral metagenomics as an emerging and powerful tool in veterinary medicine.” *The Veterinary Quart* 2011, 31(3): 107-14. doi:10.1080/01652176.2011.604971