



ANTIBIOSIS OF ENDOPHYTIC BACTERIA AGAINST PHYTOPATHOGENIC BACTERIA IN RICE CULTIVATION

Alexander Pérez Cordero^{1*}, Donicer E. Montes Vergara², Yelitza Aguas Mendoza³

¹*Universidad de Sucre, Facultad de Ciencias Agropecuarias, Colombia,
<https://orcid.org/0000-0003-3989-1747>

²Universidad de Sucre, Facultad de Ciencias Agropecuarias, Colombia,
donicer.montes@unisucra.edu.co, <https://orcid.org/0000-0002-2860-0505>

³Universidad de Sucre, Facultad de Ingeniería, Colombia, yelitza.aguas@unisucra.edu.co
<https://orcid.org/0000-0003-4880-4510>

***Corresponding Author:** Alexander Pérez Cordero

*Universidad de Sucre, Facultad de Ciencias Agropecuarias, Sincelejo, Sucre, Colombia
alexander.perez@unisucra.edu.co

Submitted: 17 November 2022.

Accepted: 21 December 2022.
jptcp (ISSN: 1710-6222)

Published: 8 February 2023

ABSTRACT

The aim of this study was to isolate endophytic bacteria from different tissues of commercial rice varieties and to test in vitro their antibacterial activity against *Burkholderia glumae* causing bacterial blast disease of rice. The results of this study report a high diversity of culturable endophytic bacteria with the ability to produce secondary metabolite type extracts with the capacity to inhibit in vitro the growth of *B. glumae*. The results obtained from the extracts at different extraction times have similar effects to that shown by the chemical control with oxolinic acid. Further studies will identify the groups of metabolites produced by these bacteria and their possible use for field management of *B. glumae*.

Keywords: bacterial extracts, inhibition, phytopathogen, rice.

INTRODUCTION

Rice (*Oryza sativa* L.) is one of the most consumed major cereals of economic interest worldwide. The production system is affected by various plant pathogens capable of causing significant economic losses in crop production and productivity (Fahad et al., 2014; Ruzmi et al., 2017; Stuart et al., 2020). Bacterial species identified as *Burkholderia glumae* and *Burkholderia gladioli* are considered the pathogenic agents of the disease known as rice panicle blast (Ham et al., 2011; Ham et al., 2018). This disease is most prevalent at the flowering stage and causes infertility, with discoloration and staining of the developing glume due to toxoflavin secretion (Lee et al., 2016).

Rice plant-associated endophytic bacteria in their process of co-evolved with the rice plant to establish themselves to the plant and reduce diseases in the shared ecological niche by employing different strategies (Ham et al., 2011). In that perspective, biological control, using native endophytic bacteria, has shown promise (Barra et al., 2014; Choi et al., 2018). Strategies based on biological mechanisms such as anti quorum sensing, quorum quenching (Fetzner, 2015), induced systemic resistance

(Chung et al., 2015) and production of antimicrobial metabolites (Ramos et al., 2019) are the most promising for disease management in the field (Shrestha et al., 2016).

Endophytic bacteria are colonizers of healthy plant tissues without symptoms of disease or intoxication, which live on the surface of clean tissues or in plant extracts (Hallmann et al., 199; Sessitsch et al., 2004). They have been isolated from a variety of plant species including rice, yams, pastures, medicinal and aromatic plants among others.

Biological control of crop diseases and pests using antagonistic microorganisms has been an environmentally friendly alternative to the use of chemical pesticides (Moenne-Loccoz et al., 2001) and is being widely studied in many plant diseases, using a diverse group of antagonistic microorganisms as part of integrated disease management programmes.

Based on the above, the strategy was to evaluate in vitro the inhibitory capacity of extracts of endophytic bacteria isolated from rice plants against *Burkholderia glumae*.

MATERIALS AND METHODS

Isolation, counting, strain separation, antibacterial activity testing of cells and metabolite compounds, strain identification by molecular sequencing and identification of the composition of microbial metabolites were performed following the scheme as described in the process in figure 1.

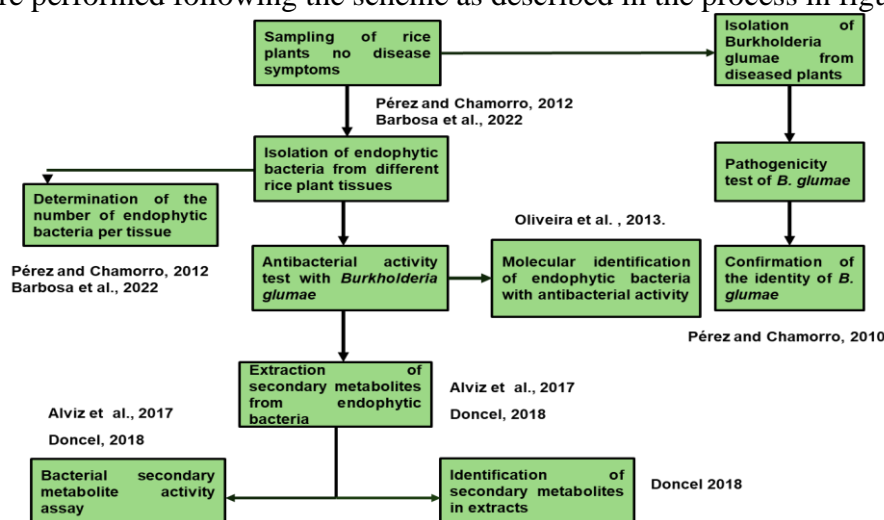


Figure 1. Stages in the process of evaluation of secondary metabolite-like compounds produced by endophytic bacteria isolated from rice plants against *Burkholderia glumae*.

RESULTS AND DISCUSSION

The sampling of rice plants was carried out at the facilities of the experimental farm La Victoria, in the locality of Mocarí, municipality of Montería, department of Córdoba, Colombia, as shown in figure 2.



Figure 2. Sampling site of commercial rice varieties for isolation of endophytic bacteria and secondary metabolite-type compounds against *Burkholderia glumae*.

A high diversity of endophytic bacterial strains was isolated from different rice plant tissues on R2A agar medium surfaces, as shown in figure 3.



Figure 3. Process of isolation, separation and purification of endophytic bacterial strains from different rice tissues.

Figure 4 shows the antibacterial activity test of microbial metabolites of endophytic bacteria against *B. glumae*. The figure shows the inhibition halo activity of the bacterial extracts compared to the positive control with oxolinic acid.

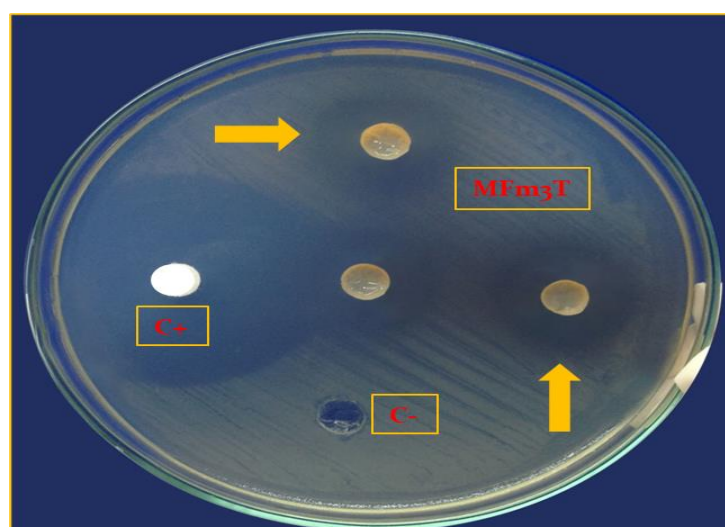


Figure 4. Antibacterial activity by production of antimicrobial compounds from endophytes bacterial isolated from rice varieties against *B. glumae*.

Figure 5 shows the antibacterial activity of endophytic bacterial cell extracts at different extraction times and the activity compared to the effect of the positive control using the chemical control with oxolinic acid.

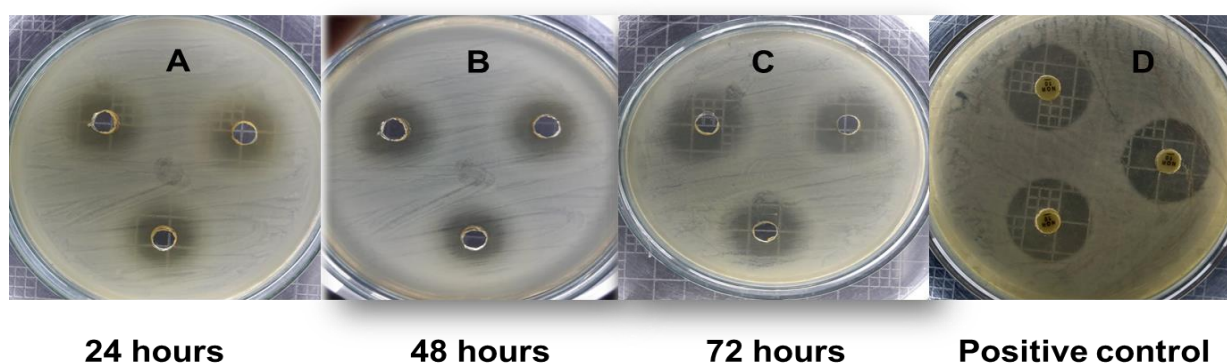


Figure 5. Antibacterial activity of crude extracts of endophytic bacteria against *B. glumae* at different exposure times.

Figure 6 shows the PCR amplification products of 16S rDNA fragment, with primers F984GC/R1378, of the total of endophytic bacteria associated with the Fmocari strain. Figure 6 shows the PCR amplification products of 16S rDNA fragment, with primers F984GC/R1378, of the total of endophytic bacteria associated with the strain F733.

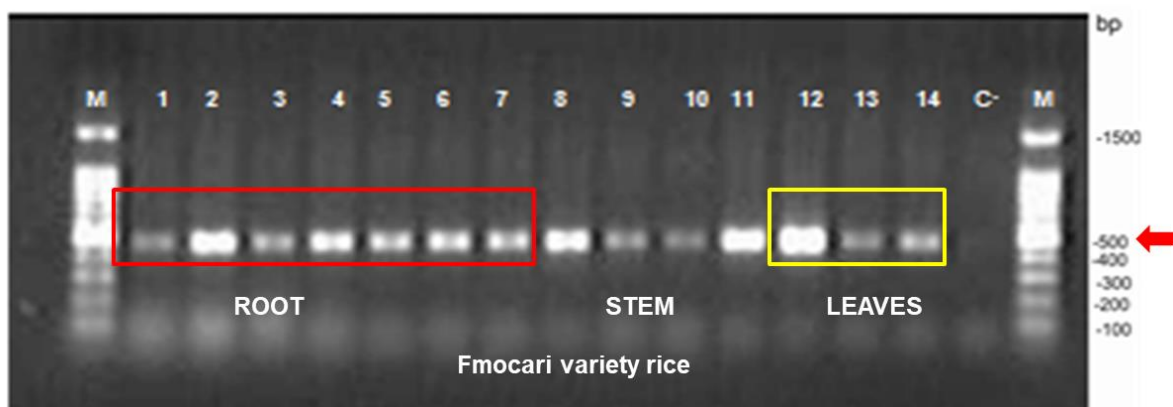


Figure 6. Agarose gel electrophoretic profile at 1.2 % of the PCR amplification product of 16S rDNA fragment, with primers F984GC/R1378, of the total of endophytic bacteria associated with the Fmocari variety. 1-7: roots, 8-11: stem, 12-14: leaf, M-100 bp DNA Ladder, C-: negative control.



Figure 7. Electrophoretic profile in agarose gel at 1.2 % of the PCR amplification product of 16S rDNA fragment, with primers F984GC/R1378, of the total of endophytic bacteria associated with variety F733. 1-7: roots, 8-11: stem, 12-14: leaf, M-100 bp DNA Ladder, C-: negative control.

Various endophytic bacteria have shown antimicrobial activity against different plant pathogens. Verma et al., 2009, conducted studies on the antimicrobial activity of endophytes associated with the neem tree, *Azadirachta indica*, in India. The endophytes were identified as *Streptomyces* sp., *Streptosporangium* sp. and *Nocardia* sp. Assays with extracts of these endophyte strains showed activity against *Pseudomonas fluorescens*, *B. subtilis*, *S. aureus*, *E. coli*, *C. albicans*, *Trichophyton* sp., *Microsporium* sp., *Aspergillus* sp., *Pythium* sp. and *Phytophthora* sp.

Similarly, Machavariani et al., (2014), isolated endophytic bacteria from *Aloe arborescens*, *Mentha arvensis*, *Lysimachia nummularia*, *Fragaria vesca* and *Arctium lappa*; native to Russia. These isolates were identified as *Nocardiopsis*, *Streptomyces* and *Micromonospora*. The results of the well diffusion assays showed activity against *S. aureus*, *Micrococcus luteus*, *B. subtilis*, *E. coli*, *P. aeruginosa* and the fungus *S. cerevisiae*. The strain extract showed activity against *E. coli*, *Salmonella* sp., *B. subtilis*, *E. faecium*, *S. aureus* and *C. albicans*. Significant disease suppression was also observed for wheat plants colonised with *B. subtilis* (Liu et al., 2009) and for banana plants pre-inoculated with endophytic *Pseudomonas* and *Burkholderia* (Fishal et al., 2010).

CONCLUSION

The results of this study show a high diversity of culturable endophytic bacteria with the ability to inhibit in vitro the growth of the phytopathogenic bacterium *Burkholderia glumae* causing bacterial blast in rice.

ACKNOWLEDGEMENTS

The authors would like to thank the microbiological research laboratory of the University of Sucre and the experimental center La Victoria of Fedearroz for their support and collaboration.

AUTHOR CONTRIBUTION. Alexander Perez Cordero: experiment execution, data analysis. Donicer Montes V and Yelitza Aguas M, conceptualization, writing - revision and editing. All authors have read and approved the manuscript.

CONFLICT OF INTEREST. All the authors of the manuscript declare that they have no conflict of interest.

REFERENCES

1. Alviz M, L., Pérez G, A., y Pérez-Cordero, A. 2017. Efecto inhibitorio de compuestos tipo metabolitos de bacterias endófitas contra *Colletotrichum gloeosporioides* y *Burkholderia glumae*. *Revista Colombiana de Ciencia Animal - RECIA*, 9(S1), 18–25. <https://doi.org/10.24188/recia.v9.nS.2017.516>
2. Barbosa-García Adrian, Pérez Cordero Alexander, Montes Vergara Donicer E. 2022. Bioprospecting Of Endophytic Bacteria With Plant Growth Promoting Activity Associated With Rice Varieties From The Colombian Caribbean. *Journal of Namibian Studies*, 32: 193-216.
3. Barraza Z, Bravo A, Pérez-Cordero A. 2014. *Pseudomonas aeruginosa* productora de metabolito con actividad antimicrobiana contra *Burkholderia glumae*. *Rev Colombiana Cienc Anim RECIA*. 114--21.
4. Choi JE, Nguyen CM, Lee B, Park JH, Oh JY, Choi JS, Song JK. 2018. Isolation and characterization of a novel metagenomic enzyme capable of degrading bacterial phytotoxin toxoflavin. *PLOS ONE*. 13:e0183893
5. Chung EJ, Hossain MT, Khan A, Kim KH, Jeon CO, Chung YR. 2015. *Bacillus oryzicola* sp. nov., an endophytic bacterium isolated from the roots of rice with antimicrobial, plant growth promoting, and systemic resistance inducing activities in rice. *Plant Pathol J*. 31:152.
6. Doncel Manrique, P. 2018. Bacterias endófitas aisladas del cultivo de ñame (*dioscorea* spp.) con producción de metabolitos con actividad antifúngica contra *colletotrichum gloeosporioides* penz. en el departamento de Sucre. Universidad de Sucre.
7. Fahad S, Nie L X, Khan F A, Chen Y T, Hussai, S, Wu C, Xiong D L, Jing W, Saud S, Khan F A, Li Y, Wu W, Khan F, Hassan S, Manan A, Jan A, Huang J L. 2014. Disease resistance in rice and the role of molecular breeding in protecting rice crops against diseases. *Biotechnol Lett*. 36: 1407–1420
8. Fetzner S. Quorum quenching enzymes. *J Biotechnol*. 2015;201:2---14
9. Hallmann J., Quadt-Hallmann A., MahaffW. F., Kloepper J.W., 1997. Bacterial endophytes in agricultural crops, *Can. J. Microbiol*. 43 (10): 895–914. <https://doi:10.1139/m97-131>.
10. Ham JH, Melanson RA, Rush MC. *Burkholderia glumae*: Next major pathogen of rice?.2011. *Mol Plant Pathol*. 12:329---39.
11. Kim J, Manna M, Kim N, Lee C, Kim J, Park J, Seo YS. 2018. The roles of two hfq genes in the virulence and stress resistance of *Burkholderia glumae*. *Plant Pathol J*. 34:412.
12. Lee HH, Park J, Kim J, Park I, Seo YS. 2016. Understanding the direction of evolution in *Burkholderia glumae* through comparative genomics. *Curr Genet*. 62:115---23
13. Liu B, Qiao H, Huang L, Buchenauer H, Han Q, Kang Z, Gong Y. 2009. Biological control of take-all in wheat by endophytic *Bacillus subtilis* E1R-j and potential mode of action. *Biol. Control* 49: 277-285.

14. Machavariani N.G., Ivankova T.D., Sineva O.N., Terek hova L.P. 2014. Isolation of endophytic actinomycetes from medicinal plants of the Moscow region, Russia, *World Appl. Sci. J.* 30 (11) 1599–1604.
15. Moenne-Loccoz, Y., Tichy, H., O'Donnel, A., Simon, R. and O'Gara, F. 2001. Impact of 2,4-Diacetylphloroglucinolproducing biocontrol strain *Pseudomonas fluorescens* F113 on intraspecific diversity of resident culturable fluorescent *Pseudomonads* associated with the roots of field-grown sugar beet seedlings. *Appl Environ Microbiol* 67, 3418– 3425.
16. Oliveira M, SantosM, Vale M, Delvaux C, Cordero P, Ferreira B Borges, A. (2013) Endophytic microbial diversity in coffee cherries of *Coffea arabica* from southeastern Brazil. *Canadian journal of microbiology*, 59(4): 221-230.
17. Pérez, C.A.; Chamorro, A.L. 2012. Bacterias endófitas: una alternativa biológica para el control de *Burkholderia glumae* en el cultivo del arroz en Colombia. *Revista Colombiana de Ciencia Animal*, 4(1):172-184
18. Ruzmi R, Ahmad-Hamdani M S, Bakar B B. 2017. Prevalence of herbicideresistant weed species in Malaysian rice fields: A review. *Weed Biol Manag.* 17: 3–16.
19. Sessitsch A., Reiter B., Berg G. 2004. Endophytic bacterial communities of field-grown potato plants and their plantgrowth- promoting and antagonistic abilities, *Can. J. Microbiol.* 50 (4): 239–249. <https://doi:10.1139/w03-118>.
20. Shrestha BK, Karki HS, Groth DE, Jungkhun N, Ham JH. 2016. Biological control activities of rice-associated *Bacillus* sp. strains against sheath blight and bacterial panicle blight of rice. *PLoS One.* 11:e0146764.
21. Stuart A M, Kong P, Then R, Flor R J, Sathya K. 2020. Tailored solutions to combat rodent pests of rice through community-based management approaches in Cambodia. *Crop Prot (in Portuguese).* 135: 1–9.
22. Suárez Moreno ZR, Vinchira-Villarraga DM, Vergara-Morales DI, Castellanos L, Ramos FA, Guarnaccia C, Moreno-Sarmiento N. 2019. Plant-growth promotion and biocontrol properties of three *Streptomyces* spp. isolates to control bacterial rice pathogens. *Front Microbiol.* 10:290
23. Verma, V. C., Gond, S. K., Kumar, A., Mishra, A., Kharwar, R. N., & Gange, A. C. 2009. Endophytic actinomycetes from *Azadirachta indica* A. Juss.: isolation, diversity, and antimicrobial activity. *Microbial ecology*, 57(4): 749-756.