



## Evaluation Of Root Canal Cleanliness After Re-Root Canal Treatment Using Two Different File System: A Scanning Electron Microscopy Study

Astha Bramhecha<sup>1</sup>, Pradeep Solete<sup>2\*</sup>, Ganesh Jeevanandan<sup>3</sup>, Delphine Priscilla Antony S<sup>4</sup>, Srujana Hemmanur<sup>5</sup>, Adimulapu Hima Sandeep<sup>6</sup>

<sup>1</sup>Department of Conservative Dentistry and Endodontics, Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technical Sciences, Saveetha University, Chennai, India

<sup>2,4</sup>Associate Professor, Department of Conservative Dentistry and Endodontics, Saveetha Dental College, Saveetha Institute of Medical and Technical Sciences, Saveetha University

<sup>3</sup>Associate Professor, Department of Pediatric and Preventive Dentistry, Saveetha Dental College, Saveetha Institute of Medical and Technical Sciences, Saveetha University

<sup>5</sup>Senior Lecturer, Department of Conservative Dentistry and Endodontics, SRM Dental College, SRM Institute of Science and Technology, Ramapuram, Chennai

<sup>6</sup>Assistant Professor, Department of Conservative dentistry and Endodontics, Saveetha Dental College and hospitals, Saveetha Institute of Medical and Technical Sciences (SIMATS), Saveetha University, Chennai-600077, Tamil Nadu, India.

\***Corresponding author:** Pradeep Solete, Associate Professor, Department of Conservative Dentistry and Endodontics, Saveetha Dental College, Saveetha Institute of Medical and Technical Sciences, Saveetha University

**Submitted: 03 March 2023; Accepted: 14 April 2023; Published: 22 May 2023**

---

### ABSTRACT

**Aim:** To evaluate the root canal cleanliness after root canal retreatment with ProTaper Universal and Solite RS rotary file system.

**Materials and Methods:** Twenty extracted single-rooted premolar teeth were selected. The root canals were enlarged with nickel-titanium (NiTi) rotary files, filled with gutta-percha and sealer, and randomly divided into the two experimental groups. The filling materials were removed with either one of the following file systems: ProTaper Universal or Solite RS. No solvent was used for removal of gutta percha. Teeth were sectioned longitudinal and assessed under the microscope.

**Results:** Both instrumentation techniques left obturating material in the canal at apical, coronal and middle third. There was no significant difference in both groups in regards to canal cleanliness at all the three levels ( $p > 0.05$ ).

**Conclusion:** Based on the findings of our in vitro study conducted on mandibular premolars with single roots, we can conclude that both instruments used in the study left filling material inside the root canal. The evaluation of the coronal, middle, and apical thirds did not reveal any significant variations in the remaining material between the instruments or the type of filling material.

**Keywords:** Gutta-percha removal, ProTaper Universal, Re-root canal treatment, Scanning electron microscopy, Solite RS3

## INTRODUCTION

Numerous factors have been linked to the lack of success in endodontic treatment. Improper removal of debris using mechanical methods, the presence of bacteria in the canals and apex, inadequate quality of sealing, excessive or insufficient filling of the root canal, and leakage in the crown area are among the frequently identified reasons for such failures. Despite the generally favorable outcomes of endodontic treatment, failures do happen in a significant proportion of cases. (1)

According to a study conducted by Kojima et al. in 2004, the success rates of root canal fillings are lower when they are either underfilled or overfilled, while the highest success rates are observed when the fillings end at the same level as the apex or within a 2 mm range. (2) Another research conducted by Akbar in 2015 revealed that the most common cause of failure in endodontic treatment was underfilling, followed by inadequate filling and overfilling. (3)

Nonsurgical endodontic retreatment aims to restore the health of periapical tissues in cases where the initial root canal treatment was ineffective or reinfection occurred due to leakage in the crown or apex. The process involves regaining access to the entire root canal system by removing the original filling, performing additional cleaning, and resealing the canal (4). The removal of gutta-percha and sealer plays a crucial role in root canal retreatment. It is important because any necrotic tissue or bacteria that might be covered by the remaining gutta-percha or sealer could potentially lead to periapical inflammation or pain (5).

Nonsurgical approaches necessitate the complete removal of filling materials from the root canal space to achieve thorough cleaning, shaping, and obturation of the root canal system (4). Various techniques employing rotary nickel-titanium (NiTi) instruments (6–8), ultrasonic instruments (9,10), heat pluggers (11,12), and manual instruments with chemical solvents (such as chloroform, eucalyptol, and orange oil) (13,14) have been proposed for removing root filling materials. Among these, rotary NiTi instruments have demonstrated effectiveness (7,15) and time efficiency (16,17) in the removal of filling materials. However, none of the existing treatment alternatives appear to ensure complete debris-free canal walls (13,16).

Introduced in 2001, the ProTaper Universal Tulsa (Dentsply Tulsa, Tulsa, OK) system, integrated with three ProTaper retreatment files (D1, D2, D3), was developed to facilitate the removal of filling materials (21, 22). The D1 PTUS instrument features an active tip, measuring 16 mm in length, 0.30 mm in tip diameter, and exhibiting a 0.09% taper. The D2 PTUS instrument has a length of 18 mm, a tip diameter of 0.25 mm, and an 0.08% taper. The D3 PTUS instrument measures 22 mm in length, 0.20 mm in tip diameter, and possesses a 0.07% taper (18).

Another recently developed system, the Solite RS3 system, offers distinct features in its retreatment files. Each file within the Solite RS3 retreatment files possesses distinct lengths, tapers, and apical tip diameters. The Solite RS1 file features a 0.30 cutting tip, for initial penetration into the filling material, an 8% taper, a length of 15 mm, and 10 mm of active blades. The Solite RS2 file, designed for middle third filling material removal, includes a 0.25 cutting tip, a 7% taper, an 18 mm length, and 13 mm of active blades, which are heat treated. The Solite RS3 file is equipped with a 0.20 non-cutting tip, a 6% taper, a length of 23 mm, and 18 mm of active blades, utilized for apical filling removal, also heat treated.

To the best of our knowledge, limited studies have investigated the application of the Solite RS3 file system retreatment files. After the removal of gutta-percha, open dentinal tubules are necessary to effectively eliminate bacteria using irrigants. However, there is currently no literature available regarding the cleanliness of dentinal tubules following retreatment, including the use of the Solite RS3 file system, using scanning electron microscopy (SEM).

Previously our team had a rich experience in working on various research projects across multiple disciplines (19–26). Now the growing trend in this area motivated us to pursue this project. Therefore, the purpose of this study was to compare the cleanliness of root canal walls after retreatment using ProTaper instruments (Dentsply Maillefer) and the Solite RS3 file system in single-rooted human teeth.

## MATERIALS AND METHODS

A total of twenty mandibular premolar teeth with single straight canals were selected for this study. Prior to the experiment, the teeth were

thoroughly cleaned using an ultrasonic scaler and rinsed with a sterile solution. Mesiodistal and buccolingual radiographs were taken to confirm the presence of a single canal. To ensure uniformity among the samples, the teeth were decoronated at the cemento-enamel junction using a diamond disc (Isomet 2000; Buehler Ltd, Lake Bluff, IL), and their root lengths were standardized to approximately 16 mm.

### **Canal Preparation**

All procedures were performed by a single operator, and teeth with calcification or multiple canals were excluded from the study. The access cavity was created using a high-speed carbide bur with water spray. After removal of the pulp tissue, a size 10 K-file (Dentsply Maillefer) was used to confirm the patency of the canal, and the working length was established at the apical foramen. The working length was set 1 mm short of the measured length. The glide path was created, and the ProTaper (Dentsply Maillefer, Ballaigues, Switzerland) files were used for cleaning and shaping with the assistance of the X-Smart Plus Endomotor (Dentsply Maillefer, OK, USA) operating at 500 rpm and 2 Ncm torque for the rotary file system. Before each file was introduced into the canal, it was lubricated with Endo Prep RC (Stedman Anabond, Chennai, India). During the preparation, 2 ml of 5% sodium hypochlorite (NaOCl) was used as an irrigant between each file. The canals were prepared until the F3 instrument. Following instrumentation, the canals were irrigated with 5 ml of 17% liquid ethylenediaminetetraacetic acid (EDTA), followed by 5 ml of saline solution. Finally, the canals were dried using paper points (Dentsply Maillefer, Ballaigues, Switzerland) corresponding to the file sizes.

### **Canal Obturation**

After drying the root canals with paper points, lateral compaction was used for obturation. A master gutta-percha cone size 30 was selected, and the tug-back was checked. AH Plus sealer (Dentsply DeTrey, Konstanz, Germany) was mixed according to the manufacturer's instructions. The master cone was coated with sealer and placed in the canal. Additional gutta-percha cones were laterally compacted until they could not be inserted more than 5 mm into the canal. Temporary filling material (Cavit, Detrey,

Dentsply) was used to seal the access cavity. The quality of the root filling was assessed by examining mesiodistal and buccolingual radiographs to ensure the absence of voids. The teeth were stored at 37°C in a humid environment for two weeks to allow complete sealer setting.

### **Retreatment Technique**

The teeth were randomly divided into two groups of ten each. The coronal filling was removed to provide access to the canal entrance. The X-Smart Plus Endomotor (Dentsply Maillefer, OK, USA) was used at the manufacturer-recommended preset torque levels for each type of instrument in the two experimental groups. In group A, ProTaper System (Dentsply Maillefer) instruments D1, D2, and D3 were used in the crown-down technique until the D3 instrument reached the working length. In group B, the gutta-percha and sealer were removed using the Solite RS3 file system in a crown-down technique, utilizing the RS1, RS2, and RS3 files in sequence until the working length was reached. No solvents were used for the removal or softening of gutta-percha in both groups.

### **Sample Analysis**

Two longitudinal grooves were made on the outer surface of the roots, and the roots were split longitudinally using a diamond disk and chisel. Care was taken to avoid entering the canal lumen with the disk. The remaining gutta-percha and sealer were evaluated in three segments: 1 mm above the apex (apical), 8 mm from the apex (middle), and 2 mm below the cemento-enamel junction (coronal). The half of each specimen with the most filling material was selected for analysis. There was no distinction made between residual sealer and gutta-percha.

### **SEM Preparation**

Selected specimens were dehydrated using graded alcohol concentrations, dried, and then coated with gold using a sputter coater (Sputter Coater; SPI, Toronto, Canada). The specimens were observed using a scanning electron microscope (JEOL 5200; JEOL, Tokyo, Japan). SEM photos of each third of the root canal (coronal, middle, and apical) were taken at a magnification of X 2000 to assess the cleanliness of the canals. The images were digitally saved

using proprietary software (SemA-fore; JEOL) and blindly evaluated by two trained operators.

**SEM Evaluation Method**

The amount of residual filling material was scored according to Ezzie *et al.*'s (27) criteria:

1. No to slight presence (0-25%) of obturation debris on the dentinal surface
2. Some presence (25-50%) of obturation debris on the dentinal surface
3. Moderate presence (50-75%) of obturation debris on the dentinal surface
4. Heavy presence (75%) of obturation debris on the dentinal surface

Each portion of the canal was divided into two fields each having a diameter of 2 to 3 mm to be evaluated and graded.

**RESULTS**

**Statistical Analysis**

The intraclass correlation coefficient was calculated to estimate the reliability of the measurements taken by the 2 examiners. The

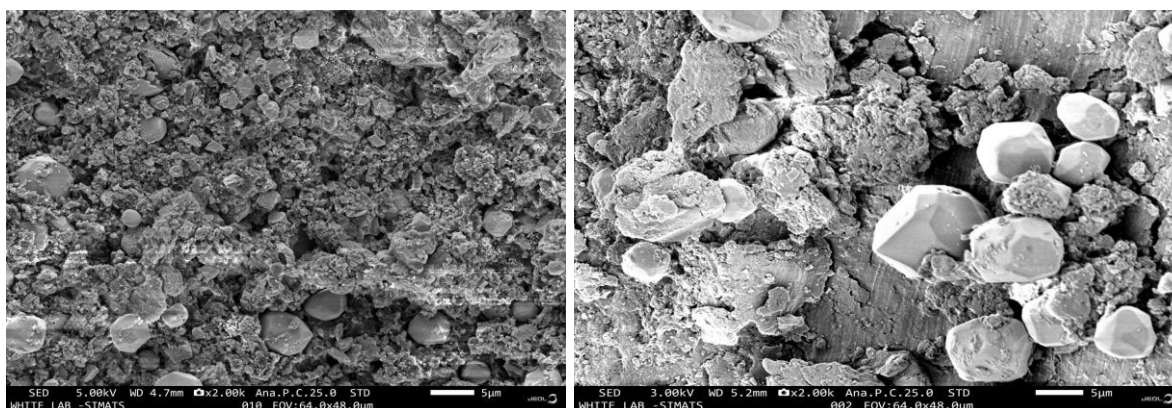
percentage of remaining filling material was evaluated for each group. Mean values for debris score were tabulated and analyzed using the statistical analysis software the Statistical Package for Social Sciences (SPSS). The chi-square test and Independent sample t-test was used to identify differences between the groups at the apical, middle, and coronal levels. The *P* value was considered statistically significant if <0.05.

**Electron Microscopy**

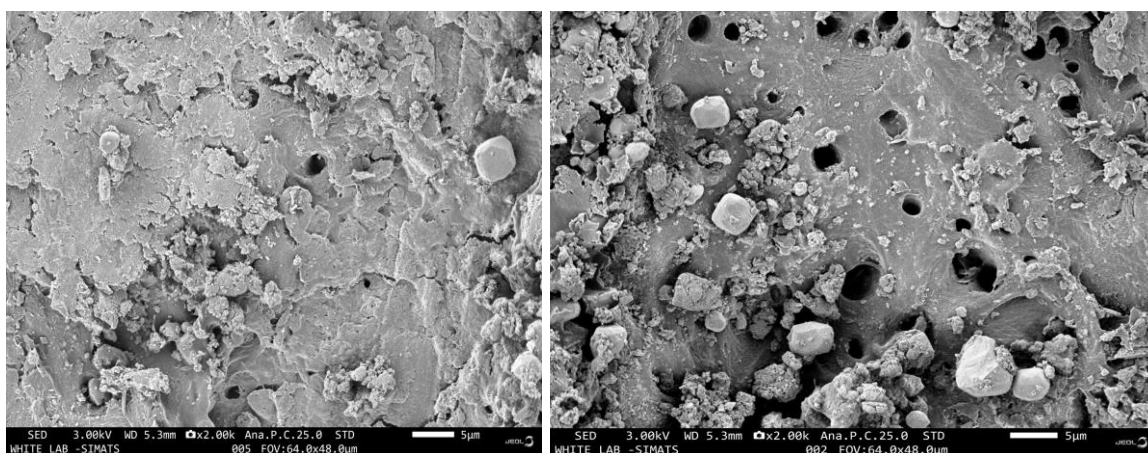
SEM of selected representative samples from each group revealed Canals in both groups tended to have increasingly more debris apically (Fig. 1) (Fig 2). In the apical area, canal walls appeared to be covered with residual obturation material and smear layer, no open dentinal tubules can be seen. Clean dentin walls and patent dentinal tubules can be observed in areas in the coronal third (Fig3). The result stated that both instrumentation techniques left obturating material with no statistically significant difference at coronal(p=0.180), middle(0.756), apical(p=0.753).

**TABLE 1:** The percentage of canals with different cleanliness scores in each treatment group

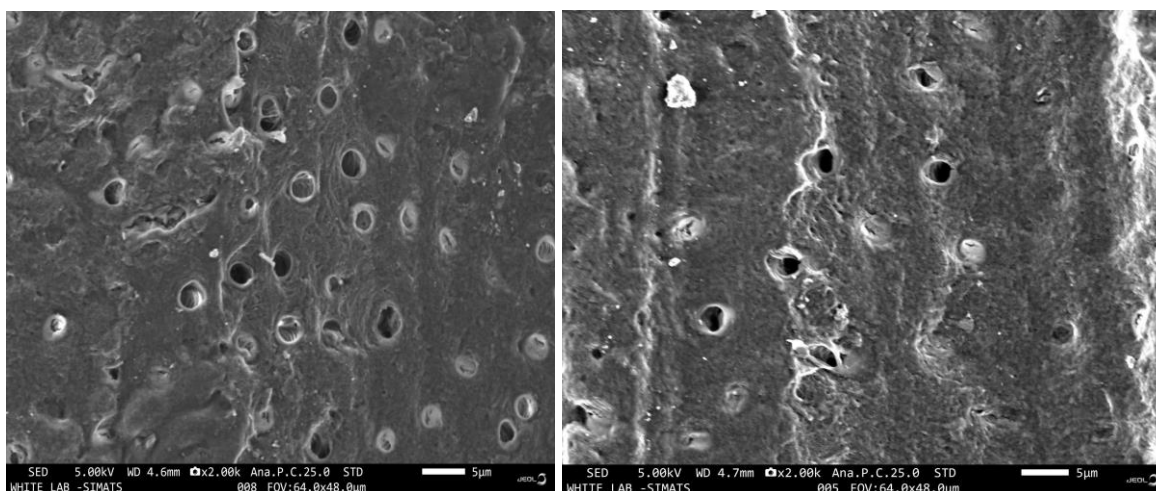
Areas	Score	Group		p-values
		ProTaper Universal	Solite RS3	
Coronal	1	35%	35%	0.18
	2	50%	40%	
	3	5%	25%	
	4	10%	0%	
Middle	1	20%	10%	0.753
	2	45%	60%	
	3	30%	25%	
	4	5%	5%	
Apical	1	10%	5%	0.756
	2	40%	50%	
	3	35%	25%	
	4	15%	20%	



**FIG 1:** Representative SEM image for Group 1(left) and Group 2(right) at the apical third showing complete obturation of dentinal tubules with obturation material.



**FIG 2:** Representative SEM image for Group 1(left) and Group 2(right) at the middle third showing partial or complete obturation of dentinal tubules with obturation material. Comparatively higher number of dentinal tubules visible.



**FIG 3:** Representative SEM image for Group 1(left) and Group 2(right) at the coronal third showing more open dentinal tubules in both the groups.

## DISCUSSION

In this particular study, the focus was on mandibular premolars, which typically have an average root length of 14 mm. However, to standardize the samples and eliminate the influence of varying lengths, the teeth were decoronated at 16 mm from the apex, leaving 2 mm of coronal structure for the coronal seal. This approach ensured consistency and comparability with previous research (27–29).

The root canals were filled using a lateral condensation technique prior to their removal, as this technique has been commonly employed in similar studies (7,30,31). The speed of the instruments used was adjusted according to the manufacturers' recommendations.

To assess the effectiveness of root filling removal, debris scores were analyzed based on Ezzie et al.'s criteria, chosen for its simplicity (27,29,32). Another scoring method provided by Hulsman and Bluhm in 2004, involving measuring gutta-percha and sealer in millimeters on a scale of 1–7, has also been used in endodontic research (6).

Various techniques have been employed in endodontic research to evaluate the efficacy of root filling removal, including radiography, digitized images, longitudinal tooth splitting, stereomicroscope visualization, digital camera imaging with software analysis, and tooth transparency methods (7,31,33–37). Each method has its advantages and limitations, providing either two-dimensional or three-dimensional information for analysis.

The results of this study indicated that none of the experimental techniques used in retreatment procedures ensured complete removal of filling materials, consistent with previous findings (7,29,38). In this study, solvent application was not employed, as recent research by Horvath et al. demonstrated that it resulted in more gutta-percha remnants on canal walls and dentinal tubules (39).

The present in vitro study employed two different rotary instruments for endodontic retreatment. Both groups of retreated teeth had relatively straight root canals initially enlarged to size 30/06, ensuring uniformity among the samples in both experimental groups. The amount of remaining filling material was evaluated through

longitudinal cleavage and quantitative analysis under SEM with 2000X magnification (28,37).

The characteristics of the cross-sectional design of Ni-Ti rotary files may contribute to variations in their cleaning ability, as observed by Hülsmann and Bluhm (6). The ProTaper Universal retreatment files remove gutta-percha in spirals around the instruments, while the Solite RS1 and RS2 files with a triple helix cross-section separate the gutta-percha from the canal walls. The ProTaper Universal files both rotate and cut the gutta-percha, whereas the Solite file System does not always cut the filling materials.

Different methods have been used in assessing canal wall cleaning, including split tooth longitudinal analysis, linear measurements of gutta-percha and sealer, microcomputer tomography, microradiographic technique, SEM, and transparent teeth methods (17,40–43). Each method has its strengths and weaknesses, providing either qualitative or quantitative evaluations.

Our institution is passionate about high quality evidence based research and has excelled in various fields.(21,24,44–54)The removal of root filling material from dentinal tubules is crucial to eliminate potential bacteria responsible for post-treatment disease and to ensure proper adaptation and adhesion of sealers and cements used for posts. SEM analysis in this study showed more open tubules in the coronal third of the root canal compared to the middle and apical thirds. These findings align with previous studies that reported higher remnants in the apical third (55,56).

Our institution is passionate about high quality evidence based research and has excelled in various fields.(57)(20–22,24,57–64).It is important to note that SEM analysis provides a more accurate assessment of dentinal tubule debris compared to macroscopic evaluation of surface remnants. The challenge in retreatment cases is that clinicians often rely on visual and radiographic analysis to evaluate the thoroughness of canal cleansing(30). In this study, a magnification of 2000X was consistently used for all SEM images, and the results were evaluated considering the specific part of the root from which the images were taken. Operator bias was minimized by grooves in the root surface indicating the designated investigation areas.

## CONCLUSION

Based on the findings of our in vitro study conducted on mandibular premolars with single roots, we can conclude that both instruments used in the study left filling material inside the root canal. The evaluation of the coronal, middle, and apical thirds did not reveal any significant variations in the remaining material between the instruments or the type of filling material.

## REFERENCES

1. Tabassum S, Khan FR. Failure of endodontic treatment: The usual suspects. *Eur J Dent* [Internet]. 2016 Jan-Mar;10(1):144–7. Available from: <http://dx.doi.org/10.4103/1305-7456.175682>
2. Kojima K, Inamoto K, Nagamatsu K, Hara A, Nakata K, Morita I, et al. Success rate of endodontic treatment of teeth with vital and nonvital pulps. A meta-analysis. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* [Internet]. 2004 Jan;97(1):95–9. Available from: <http://dx.doi.org/10.1016/j.tripleo.2003.07.006>
3. Akbar I. Radiographic study of the problems and failures of endodontic treatment. *Int J Health Sci* [Internet]. 2015 Apr;9(2):111–8. Available from: <https://www.ncbi.nlm.nih.gov/pubmed/26309429>
4. Stabholz A, Friedman S. Endodontic retreatment--case selection and technique. Part 2: Treatment planning for retreatment. *J Endod* [Internet]. 1988 Dec;14(12):607–14. Available from: [http://dx.doi.org/10.1016/S0099-2399\(88\)80058-X](http://dx.doi.org/10.1016/S0099-2399(88)80058-X)
5. Ruddle CJ. Nonsurgical retreatment. *J Endod* [Internet]. 2004 Dec;30(12):827–45. Available from: <http://dx.doi.org/10.1097/01.don.0000145033.15701.2d>
6. Hülsmann M, Bluhm V. Efficacy, cleaning ability and safety of different rotary NiTi instruments in root canal retreatment. *Int Endod J* [Internet]. 2004 Jul;37(7):468–76. Available from: <http://dx.doi.org/10.1111/j.1365-2591.2004.00823.x>
7. Schirrmeister JF, Wrbas KT, Meyer KM, Altenburger MJ, Hellwig E. Efficacy of different rotary instruments for gutta-percha removal in root canal retreatment. *J Endod* [Internet]. 2006 May;32(5):469–72. Available from: <http://dx.doi.org/10.1016/j.joen.2005.10.052>
8. Hülsmann M, Stotz S. Efficacy, cleaning ability and safety of different devices for gutta-percha removal in root canal retreatment. *Int Endod J* [Internet]. 1997 Jul;30(4):227–33. Available from: <https://onlinelibrary.wiley.com/doi/abs/10.1046/j.1365-2591.1997.00036.x>
9. Krell KV, Neo J. The use of ultrasonic endodontic instrumentation in the re-treatment of a paste-filled endodontic tooth. *Oral Surg Oral Med Oral Pathol* [Internet]. 1985 Jul;60(1):100–2. Available from: [http://dx.doi.org/10.1016/0030-4220\(85\)90222-1](http://dx.doi.org/10.1016/0030-4220(85)90222-1)
10. Jeng HW, ElDeeb ME. Removal of hard paste fillings from the root canal by ultrasonic instrumentation. *J Endod* [Internet]. 1987 Jun;13(6):295–8. Available from: [http://dx.doi.org/10.1016/S0099-2399\(87\)80047-X](http://dx.doi.org/10.1016/S0099-2399(87)80047-X)
11. Friedman S, Stabholz A, Tamse A. Endodontic retreatment—Case selection and technique. Part 3. Retreatment techniques. *J Endod* [Internet]. 1990 Nov 1;16(11):543–9. Available from: <https://www.sciencedirect.com/science/article/pii/S0099239907802196>
12. Friedman S, Mor C. The success of endodontic therapy--healing and functionality. *J Calif Dent Assoc* [Internet]. 2004 Jun;32(6):493–503. Available from: <https://www.ncbi.nlm.nih.gov/pubmed/15344440>
13. Schirrmeister JF, Wrbas KT, Schneider FH, Altenburger MJ, Hellwig E. Effectiveness of a hand file and three nickel-titanium rotary instruments for removing gutta-percha in curved root canals during retreatment. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* [Internet]. 2006 Apr;101(4):542–7. Available from: <http://dx.doi.org/10.1016/j.tripleo.2005.03.003>
14. Friedman S, Moshonov J, Trope M. Efficacy of removing glass ionomer cement, zinc oxide eugenol, and epoxy resin sealers from retreated root canals. *Oral Surg Oral Med Oral Pathol* [Internet]. 1992 May;73(5):609–12. Available from: [http://dx.doi.org/10.1016/0030-4220\(92\)90108-3](http://dx.doi.org/10.1016/0030-4220(92)90108-3)
15. Betti LV, Bramante CM. Quantec SC rotary instruments versus hand files for gutta-percha removal in root canal retreatment. *Int Endod J* [Internet]. 2001 Oct;34(7):514–9. Available from: <http://dx.doi.org/10.1046/j.1365-2591.2001.00424.x>
16. Imura N, Kato AS, Hata GI, Uemura M, Toda T, Weine F. A comparison of the relative efficacies of four hand and rotary instrumentation techniques during endodontic retreatment. *Int Endod J* [Internet]. 2000 Jul;33(4):361–6. Available from: <http://dx.doi.org/10.1046/j.1365-2591.2000.00320.x>
17. Ferreira JJ, Rhodes JS, Ford TR. The efficacy of gutta-percha removal using ProFiles. *Int Endod J* [Internet]. 2001 Jun;34(4):267–74. Available from: <https://onlinelibrary.wiley.com/doi/abs/10.1046/j.1365-2591.2000.00320.x>

- from:  
<https://onlinelibrary.wiley.com/doi/abs/10.1046/j.1365-2591.2001.00379.x>
18. Huang X, Ling J, Wei X, Gu L. Quantitative evaluation of debris extruded apically by using ProTaper Universal Tulsa rotary system in endodontic retreatment. *J Endod* [Internet]. 2007 Sep;33(9):1102–5. Available from: <http://dx.doi.org/10.1016/j.joen.2007.05.019>
  19. Poornima P, Krithikadatta J, Ponraj RR, Velmurugan N, Kishen A. Biofilm formation following chitosan-based varnish or chlorhexidine-fluoride varnish application in patients undergoing fixed orthodontic treatment: a double blinded randomised controlled trial. *BMC Oral Health* [Internet]. 2021 Sep 23;21(1):465. Available from: <http://dx.doi.org/10.1186/s12903-021-01805-8>
  20. Murugesan A, Sivakumar A. Comparison of accuracy of mesiodistal tooth measurements made in conventional study models and digital models obtained from intraoral scan and desktop scan of study models. *J Orthod* [Internet]. 2020 Jun;47(2):149–55. Available from: <http://dx.doi.org/10.1177/1465312520910755>
  21. Kurniawan A, Chusida A 'nisaa, Atika N, Gianosa TK, Solikhin MD, Margaretha MS, et al. The Applicable Dental Age Estimation Methods for Children and Adolescents in Indonesia. *Int J Dent* [Internet]. 2022 Feb 15;2022:6761476. Available from: <http://dx.doi.org/10.1155/2022/6761476>
  22. Jeevanandan G, Ravindran V, Subramanian EM, Kumar AS. Postoperative Pain with Hand, Reciprocating, and Rotary Instrumentation Techniques after Root Canal Preparation in Primary Molars: A Randomized Clinical Trial. *Int J Clin Pediatr Dent* [Internet]. 2020 Jan-Feb;13(1):21–6. Available from: <http://dx.doi.org/10.5005/jp-journals-10005-1709>
  23. Paulraj J, Nagar P. Antimicrobial Efficacy of Triphala and Propolis-modified Glass Ionomer Cement: An In Vitro Study. *Int J Clin Pediatr Dent* [Internet]. 2020 Sep-Oct;13(5):457–62. Available from: <http://dx.doi.org/10.5005/jp-journals-10005-1806>
  24. Sekar D, Murthykumar K, Ganapathy D. miR-206 and its mimics: A predictive biomarker and therapeutic molecule in the treatment of oral cancer. *Oral Oncol* [Internet]. 2022 May;128:105849. Available from: <http://dx.doi.org/10.1016/j.oraloncology.2022.105849>
  25. Harsha L, Subramanian AK. Comparative Assessment of pH and Degree of Surface Roughness of Enamel When Etched with Five Commercially Available Etchants: An In Vitro Study. *J Contemp Dent Pract* [Internet]. 2022 Feb 1;23(2):181–5. Available from: <https://www.ncbi.nlm.nih.gov/pubmed/35748447>
  26. Hegde MN, Attavar SH, Shetty N, Hegde ND, Hegde NN. Saliva as a biomarker for dental caries: A systematic review. *J Conserv Dent* [Internet]. 2019 Jan-Feb;22(1):2–6. Available from: [http://dx.doi.org/10.4103/JCD.JCD\\_531\\_18](http://dx.doi.org/10.4103/JCD.JCD_531_18)
  27. Ezzie E, Fleury A, Solomon E, Spears R, He J. Efficacy of retreatment techniques for a resin-based root canal obturation material. *J Endod* [Internet]. 2006 Apr;32(4):341–4. Available from: <http://dx.doi.org/10.1016/j.joen.2005.09.010>
  28. Só MVR, Saran C, Magro ML, Vier-Pelisser FV, Munhoz M. Efficacy of ProTaper retreatment system in root canals filled with gutta-percha and two endodontic sealers. *J Endod* [Internet]. 2008 Oct;34(10):1223–5. Available from: <http://dx.doi.org/10.1016/j.joen.2008.07.020>
  29. Somma F, Cammarota G, Plotino G, Grande NM, Pameijer CH. The effectiveness of manual and mechanical instrumentation for the retreatment of three different root canal filling materials. *J Endod* [Internet]. 2008 Apr;34(4):466–9. Available from: <http://dx.doi.org/10.1016/j.joen.2008.02.008>
  30. Cunha RS, De Martin AS, Barros PP, da Silva FM, Jacinto R de C, Bueno CE da S. In vitro evaluation of the cleansing working time and analysis of the amount of gutta-percha or Resilon remnants in the root canal walls after instrumentation for endodontic retreatment. *J Endod* [Internet]. 2007 Dec;33(12):1426–8. Available from: <http://dx.doi.org/10.1016/j.joen.2007.07.004>
  31. de Oliveira DP, Barbizam JVB, Trope M, Teixeira FB. Comparison between gutta-percha and resilon removal using two different techniques in endodontic retreatment. *J Endod* [Internet]. 2006 Apr;32(4):362–4. Available from: <http://dx.doi.org/10.1016/j.joen.2005.12.006>
  32. Marfisi K, Mercade M, Plotino G, Duran-Sindreu F, Bueno R, Roig M. Efficacy of three different rotary files to remove gutta-percha and Resilon from root canals. *Int Endod J* [Internet]. 2010 Nov;43(11):1022–8. Available from: <http://dx.doi.org/10.1111/j.1365-2591.2010.01758.x>
  33. Masiero AV, Barletta FB. Effectiveness of different techniques for removing gutta-percha during retreatment. *Int Endod J* [Internet]. 2005 Jan;38(1):2–7. Available from: <http://dx.doi.org/10.1111/j.1365-2591.2004.00878.x>
  34. de Carvalho Maciel AC, Zaccaro Scelza MF.



- Efficacy of automated versus hand instrumentation during root canal retreatment: an ex vivo study. *Int Endod J* [Internet]. 2006 Oct;39(10):779–84. Available from: <http://dx.doi.org/10.1111/j.1365-2591.2006.01148.x>
35. Unal GC, Kaya BU, Taç AG, Keçeci AD. A comparison of the efficacy of conventional and new retreatment instruments to remove gutta-percha in curved root canals: an ex vivo study. *Int Endod J* [Internet]. 2009 Apr;42(4):344–50. Available from: <http://dx.doi.org/10.1111/j.1365-2591.2008.01518.x>
36. Taşdemir T, Yildirim T, Celik D. Comparative study of removal of current endodontic fillings. *J Endod* [Internet]. 2008 Mar;34(3):326–9. Available from: <http://dx.doi.org/10.1016/j.joen.2007.12.022>
37. Giuliani V, Cocchetti R, Pagavino G. Efficacy of ProTaper universal retreatment files in removing filling materials during root canal retreatment. *J Endod* [Internet]. 2008 Nov;34(11):1381–4. Available from: <http://dx.doi.org/10.1016/j.joen.2008.08.002>
38. Hammad M, Qualtrough A, Silikas N. Three-dimensional evaluation of effectiveness of hand and rotary instrumentation for retreatment of canals filled with different materials. *J Endod* [Internet]. 2008 Nov;34(11):1370–3. Available from: <http://dx.doi.org/10.1016/j.joen.2008.07.024>
39. Horvath SD, Altenburger MJ, Naumann M, Wolkewitz M, Schirrmeyer JF. Cleanliness of dentinal tubules following gutta-percha removal with and without solvents: a scanning electron microscopic study. *Int Endod J* [Internet]. 2009 Nov;42(11):1032–8. Available from: <http://dx.doi.org/10.1111/j.1365-2591.2009.01616.x>
40. Taşdemir T, Er K, Yildirim T. Efficacy of three rotary NiTi instruments in removing gutta-percha from root canals. *Int Endod J* [Internet]. 2008; Available from: <https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1365-2591.2007.01335.x>
41. Gu LS, Ling JQ, Wei X, Huang XY. Efficacy of ProTaper Universal rotary retreatment system for gutta-percha removal from root canals. *Int Endod J* [Internet]. 2008 Apr;41(4):288–95. Available from: <http://dx.doi.org/10.1111/j.1365-2591.2007.01350.x>
42. Rhodes JS, Ford TR, Lynch JA, Liepins PJ, Curtis RV. Micro-computed tomography: a new tool for experimental endodontology. *Int Endod J* [Internet]. 1999 May;32(3):165–70. Available from: <http://dx.doi.org/10.1046/j.1365-2591.1999.00204.x>
43. Saad AY, Al-Hadlaq SM, Al-Katheeri NH. Efficacy of two rotary NiTi instruments in the removal of Gutta-Percha during root canal retreatment. *J Endod* [Internet]. 2007 Jan;33(1):38–41. Available from: <http://dx.doi.org/10.1016/j.joen.2006.08.012>
44. Arumugam P, George R, Jayaseelan VP. Aberrations of m6A regulators are associated with tumorigenesis and metastasis in head and neck squamous cell carcinoma. *Arch Oral Biol* [Internet]. 2021 Feb;122:105030. Available from: <http://dx.doi.org/10.1016/j.archoralbio.2020.10.5030>
45. Balachander K, Paramasivam A. Cell-free mitochondrial DNA as a novel non-invasive biomarker for oral cancer. *Oral Oncol* [Internet]. 2022 Apr;127:105825. Available from: <http://dx.doi.org/10.1016/j.oraloncology.2022.105825>
46. Bijai LK, Muthukrishnan A. Potential role of fibroblast senescence in malignant transformation of oral submucous fibrosis. *Oral Oncol* [Internet]. 2022 Apr;127:105810. Available from: <http://dx.doi.org/10.1016/j.oraloncology.2022.105810>
47. Ezhilarasan D, Lakshmi T, Subha M, Deepak Nallasamy V, Raghunandhakumar S. The ambiguous role of sirtuins in head and neck squamous cell carcinoma. *Oral Dis* [Internet]. 2022 Apr;28(3):559–67. Available from: <http://dx.doi.org/10.1111/odi.13798>
48. Pandiar D, Ramani P, Krishnan RP, Monica K. Multifaceted multinucleated giant cells in oral squamous cell carcinoma. *Oral Oncol* [Internet]. 2021 Oct;121:105400. Available from: <http://dx.doi.org/10.1016/j.oraloncology.2021.105400>
49. Sekar D. Circulatory microRNAs inhibition and its signaling pathways in the treatment of oral squamous cell carcinoma (OSCC). *Oral Oncol* [Internet]. 2022 Mar;126:105763. Available from: <http://dx.doi.org/10.1016/j.oraloncology.2022.105763>
50. Murugesan A, Sivakumar A. Comparison of accuracy of mesiodistal tooth measurements made in conventional study models and digital models obtained from intraoral scan and desktop scan of study models. *J Orthod* [Internet]. 2020 Jun;47(2):149–55. Available from: <http://dx.doi.org/10.1177/1465312520910755>
51. Sekar D. The Role of microRNAs as a predictive biomarker and therapeutic molecule in the treatment of Oral Potentially Malignant Disorder (OPMD). *Oral Oncol* [Internet]. 2022 Apr;127:105786. Available from: <http://dx.doi.org/10.1016/j.oraloncology.2022.105786>

- 05786
52. Jeevanandan G, Ravindran V, Subramanian EM, Kumar AS. Postoperative Pain with Hand, Reciprocating, and Rotary Instrumentation Techniques after Root Canal Preparation in Primary Molars: A Randomized Clinical Trial. *Int J Clin Pediatr Dent* [Internet]. 2020 Jan-Feb;13(1):21–6. Available from: <http://dx.doi.org/10.5005/jp-journals-10005-1709>
  53. Preethy NA, Jeevanandan G, Govindaraju L, Subramanian E. Comparison of Shear Bond Strength of Three Commercially Available Esthetic Restorative Composite Materials: An Study. *Int J Clin Pediatr Dent* [Internet]. 2020 Nov-Dec;13(6):635–9. Available from: <http://dx.doi.org/10.5005/jp-journals-10005-1849>
  54. Govindaraju L, Jeevanandan G, Subramanian EMG. ABO blood grouping: A potential risk factor for early childhood caries - A cross-sectional study. *Indian J Dent Res* [Internet]. 2018 May-Jun;29(3):313–6. Available from: [http://dx.doi.org/10.4103/ijdr.IJDR\\_156\\_17](http://dx.doi.org/10.4103/ijdr.IJDR_156_17)
  55. Wilcox LR. Endodontic retreatment: ultrasonics and chloroform as the final step in reinstrumentation. *J Endod* [Internet]. 1989 Mar;15(3):125–8. Available from: [http://dx.doi.org/10.1016/S0099-2399\(89\)80133-5](http://dx.doi.org/10.1016/S0099-2399(89)80133-5)
  56. Gergi R, Sabbagh C. Effectiveness of two nickel-titanium rotary instruments and a hand file for removing gutta-percha in severely curved root canals during retreatment: an ex vivo study. *Int Endod J* [Internet]. 2007 Jul;40(7):532–7. Available from: <https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1365-2591.2007.01254.x>
  57. Sekar D. Circulatory microRNAs inhibition and its signaling pathways in the treatment of oral squamous cell carcinoma (OSCC). *Oral Oncol* [Internet]. 2022 Mar;126:105763. Available from: <http://dx.doi.org/10.1016/j.oraloncology.2022.105763>
  58. Arumugam P, George R, Jayaseelan VP. Aberrations of m6A regulators are associated with tumorigenesis and metastasis in head and neck squamous cell carcinoma. *Arch Oral Biol* [Internet]. 2021 Feb;122:105030. Available from: <http://dx.doi.org/10.1016/j.archoralbio.2020.105030>
  59. Balachander K, Paramasivam A. Cell-free mitochondrial DNA as a novel non-invasive biomarker for oral cancer. *Oral Oncol* [Internet]. 2022 Apr;127:105825. Available from: <http://dx.doi.org/10.1016/j.oraloncology.2022.105825>
  60. Bijai LK, Muthukrishnan A. Potential role of fibroblast senescence in malignant transformation of oral submucous fibrosis. *Oral Oncol* [Internet]. 2022 Apr;127:105810. Available from: <http://dx.doi.org/10.1016/j.oraloncology.2022.105810>
  61. Ezhilarasan D, Lakshmi T, Subha M, Deepak Nallasamy V, Raghunandhakumar S. The ambiguous role of sirtuins in head and neck squamous cell carcinoma. *Oral Dis* [Internet]. 2022 Apr;28(3):559–67. Available from: <http://dx.doi.org/10.1111/odi.13798>
  62. Pandiar D, Ramani P, Krishnan RP, Monica K. Multifaceted multinucleated giant cells in oral squamous cell carcinoma. *Oral Oncol* [Internet]. 2021 Oct;121:105400. Available from: <http://dx.doi.org/10.1016/j.oraloncology.2021.105400>
  63. Sekar D. The Role of microRNAs as a predictive biomarker and therapeutic molecule in the treatment of Oral Potentially Malignant Disorder (OPMD). *Oral Oncol* [Internet]. 2022 Apr;127:105786. Available from: <http://dx.doi.org/10.1016/j.oraloncology.2022.105786>
  64. Preethy NA, Jeevanandan G, Govindaraju L, Subramanian E. Comparison of Shear Bond Strength of Three Commercially Available Esthetic Restorative Composite Materials: An Study. *Int J Clin Pediatr Dent* [Internet]. 2020 Nov-Dec;13(6):635–9. Available from: <http://dx.doi.org/10.5005/jp-journals-10005-1849>