



Antimicrobial property and Cytotoxicity of hesperidin incorporated dentin adhesive – an invitro study

M. Shamly¹, Iffat Nasim^{2*}

¹Post graduate student, Department of Conservative dentistry and Endodontics, Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technical Sciences, Saveetha University

²Head of the Department, Department of Conservative dentistry and Endodontics, Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technical Sciences, Saveetha University

***Corresponding author:** Iffat Nasim, Head of the Department, Department of Conservative dentistry and Endodontics, Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technical Sciences, Saveetha University, Email: iffatnasim@saveetha.com

Submitted: 21 March 2023; Accepted: 18 April 2023; Published: 06 May 2023

ABSTRACT

Introduction: Dentin adhesives are frequently used in restorative dentistry, although they can be vulnerable to bacterial colonisation, which can result in recurrent caries. Citrus fruits contain the flavonoid hesperidin (HPN), which has demonstrated potential antibacterial activity against a variety of microorganisms. This study looked at the cytotoxicity and antibacterial effects of total etch dentin adhesive that was incorporated with hesperidin.

Materials and Methods: Four groups of dentin adhesive formulations with different hesperidin concentrations (25µL, 50µL, and 100µL) were made. The test microbes for the antimicrobial investigation were *Lactobacillus acidophilus*, *Streptococcus mutans*, and *Enterococcus faecalis*. For this experiment, Mueller Hinton Agar was used to measure the zone of inhibition. Also, the cytotoxicity of the hesperidin-incorporated total etch dentin adhesive was examined using the Brine Shrimp Lethality Assay.

Results: Dimethyl sulfoxide (DMSO), total etch bonding agent, and HPN work together synergistically to combat *L. acidophilus*, *E. faecalis*, and *S. mutans* in particular. Hesperidin concentration in the adhesive led to an expansion of the zone of inhibition. The hesperidin-incorporated adhesive did not have any harmful effects, as shown by the Brine shrimp lethality experiment.

Conclusion: According to the results of this in vitro investigation, adding hesperidin to dentin adhesive may improve its antibacterial capabilities without harming dental pulp stem cells. These findings open up new directions for investigation into the development of dental adhesive formulations with enhanced antibacterial characteristics for superior clinical results.

Clinical Significance: Hesperidin and other natural substances could be used into dentin adhesive to prevent bacterial colonisation and avoid recurrent caries. Furthermore, the hesperidin-incorporated adhesive's lack of cytotoxicity proves that it is safe for usage in clinical settings.

Keywords: *Hesperidin, Flavonoid, Total etch dentin adhesive, Micro-organisms, Quality of life*

INTRODUCTION

Dental caries is a prevalent oral health issue that affects people of all ages worldwide. Dentin adhesive agents are commonly used in restorative dentistry to establish strong bonds between the tooth structure and restorative materials¹. However, these adhesives are susceptible to bacterial colonization, leading to recurrent caries and treatment failures. The dentin-resin hybrid layer is regarded as the weak link in resin-based restorations². Unpolymerised resins may affect the hybrid layer, may be sensitive to dentin and may alter the pulpal physiology. Flavonoids acts as a natural collagen cross linker in dentin. Incorporating natural compounds, such as flavonoids, into dentin adhesive formulations has emerged as a promising approach to improving their antimicrobial properties and reducing the risk of recurrent caries³. Total etch dentin adhesives are preferred because they are proved to have considerable bond strength compared to self etch dentin adhesives⁴. However, there are certain drawbacks, like collagen deterioration from acid etching⁵. MMPs and cysteine cathepsins, called pro enzymes that are produced from the host, are inactive⁶. They become active at lower pH levels as a result of acid conditioning, which can lead to the destruction of collagen, elastin, and extracellular matrix (ECM)⁷. This has an impact on the hybrid layer and reduces stability of the composite resin's bonds⁸. Furthermore, etching gelatinizes collagen fibres and limits resin diffusion in interfibrillar regions. Collagen fibres that aren't protected can then degrade. Its degradation can be avoided by adding substances with collagen crosslinking and MMP inhibitory properties to total etch⁹. A class of plant substances called flavonoids are well known for being anti-inflammatory and antioxidant. It has been demonstrated that they may be utilised to enhance the qualities of dental materials, particularly adhesives used in restorative dentistry.

Hesperidin, a flavonoid compound found in citrus fruits, has received considerable attention for its potential health-promoting properties¹⁰. Recent studies have highlighted the antimicrobial activity of flavonoids against various bacteria, including *Streptococcus mutans* and *Lactobacillus acidophilus*, which are the primary

causative agents of dental caries^{11,12}. Hesperidin possesses anti-inflammatory and antioxidant properties, making it an ideal candidate for use in dental adhesives. The incorporation of hesperidin into dentin adhesive may show promising results in improving its antimicrobial properties^{13,14}. Several studies have reported that the addition of Hesperidin has minimal cytotoxic effects on human cells, making it a safe and effective additive for dental adhesives.

The objective of this study was to investigate the antimicrobial property and cytotoxicity of hesperidin-incorporated total etch dentin adhesive using in vitro assays. Specifically, we aimed to evaluate the effect of varying concentrations of hesperidin (25µL, 50 µL, 100 µL) on bacterial growth and dental pulp stem cell viability. The findings of this study may provide valuable insights into the development of new dental adhesive formulations with enhanced antimicrobial properties for better clinical outcomes. The incorporation of natural compounds, such as hesperidin, into dentin adhesive could provide a safe and effective way to combat bacterial colonization and prevent recurrent caries. The results of this study may pave the way for further research to optimize the concentration and delivery of hesperidin in dental adhesive formulations to improve the quality of life by providing long lasting restorations. The null hypothesis states that there is no discernible difference between commercial dentin adhesive and dentin adhesive with hesperidin. Our team has extensive knowledge and research experience that has translate into high quality publications^{15–24,25–29}

MATERIALS & METHODS

Preparation of test solution

2% of Hesperidin (HPN) is incorporated into dentin adhesive (2mg powder in 98ml of bonding agent). Adper single bond 2 was the total etch adhesive used in this study. 2 % HPN provides immediate bond strength and does not produce discoloration. Dimethyl sulphoxide was used in little amount as a solvent to solubilize hesperidin. Hesperidin powder (Sigma–Aldrich) was directly dissolved into pure Dimethyl sulfoxide (20mg of HPN in 0.025ml of

DMSO).The parent material was Adper Single Bond 2, a total etch dentin adhesive that is sold over the counter (3 M ESPE). Then the Hesperidin/Dimethyl sulfoxide was incorporated into Adper single bond 2 at proper ratio (20mg of HPN in 1 ml of bonding agent) to get the final concentration 2% hesperidin in the total etch adhesive employed.

Control group: Flavonoid(HPN)free adhesive

Test group: Flavonoid(HPN) incorporated adhesive (20mg HPN+0.025ml DMSO+ 1ml of Adper single bond 2)

Antimicrobial activity

In order to identify the zone of inhibition, Mueller Hinton Agar was used.

o The test organisms were swabbed after the wells were cut with a 9 mm sterile polystyrene tip.



Brine Shrimp Lethality Assay

RESULTS

Measuring the diameter of the zone of inhibition surrounding the substance that was tested against the target microbiological organism is necessary for interpreting the zone of inhibition assay, a technique used to evaluate a substance's antimicrobial efficacy. The zone of inhibition is the region around the substance where the microbe's growth has been impeded or ceased. The effectiveness of the chemical against the microorganism increases with the diameter of the zone of inhibition. The type of organism being tested, the chemical being utilised, and the substance's concentration can all affect the

o Several concentrations of the experimental adhesive (25, 50, and 100 mL) were loaded, and standard antibiotic amoxyrite was injected in the fourth well.

The plates were incubated at 37 °C for 24 hours. The zone of inhibition was assessed following the incubation period.

Assessment of cytotoxicity

o Ten to twelve millilitres of saline water were added to six-well ELISA plates. A total of 10 nauplii (5, 10, 20, 40, 80, and control-normal saline) were gradually added to each well.

Thereafter, in accordance with the concentration level, the experimental adhesive solutions were added. 24 hours were spent incubating the plates.

o The ELISA plates were examined after 24 hours and counted for the presence of live nauplii.

breakpoints for interpreting the zone of inhibition assay.

With increasing hesperidin-incorporated dentin adhesive concentration, the zone of inhibition expands. The most frequent cariogenic microbes, lactobacillus acidophillus and streptococcus mutans, show the greatest inhibition. Enterococcus faecalis exhibits significantly reduced inhibition. The following figures (a,b,c) and table(1) show the outcomes.



Zone of inhibition in relation to Lactobacillus acidophillus



Zone of inhibition in relation to *Enterococcus faecalis*



Zone of inhibition in relation to *Streptococcus mutans*

TABLE 1: Measurement of Zone of inhibition of test and control groups

	Control	Test (25µL)	Test (50µL)	Test (100µL)
L.acidophilus	13mm	14mm	16mm	18.5mm
E.faecalis	9mm	10mm	12mm	14mm
S.mutans	10mm	11mm	12mm	13.5mm

The bioassay known as the brine shrimp lethality assay is frequently used to assess the toxicity of chemical substances. The dead shrimp in each concentration of the substance under test is counted in order to evaluate the assay's results. The relationship between a chemical's concentration and a response is known as a dose-response relationship (in this case, shrimp

mortality). The toxicity of the drug and the success of the assay can both be determined using this connection. The concentration of a substance at which 50% of shrimp die is known as the LC50 (lethal concentration 50). The higher LC50 in this investigation indicates reduced cytotoxicity. Table 2 presents the findings.

TABLE 2: Assessment of live nauplii's in relation to concentration of hesperidin incorporated into dentin adhesive

Concentration (µL)	No. of live nauplii's
5	9
10	8
20	7
40	7
80	7
Control	10

DISCUSSION

Flavonoids have antimicrobial properties, collagen-crosslinking effects, and dentin collagen proteolytic degradation inhibition³⁰. The mechanical properties of dentine are improved by the cross-linking of collagen and other proline-rich proteins by a naturally occurring flavonoid called Hesperidin. Yet, depending on the chemical composition of the flavonoids and the type of microbe, different flavonoids have different antibacterial potencies³¹. When exposed to flavonoids like hesperidin, several bacteria become immune to infection and multiplication³². Molecular structure, hydrophobicity, solubility, the presence or absence of a sugar moiety, and the kind of sugar in the chemical backbone all have an impact on the antibacterial activity and bioavailability of flavonoids³³. Yet, it is still unknown exactly how flavonoids work to prevent bacteria from growing. The interruption of bacterial DNA synthesis, bacterial motility, cytoplasmic membrane permeability, and inhibition of bacterial metalloenzymes are only a few of the methods that have been put forth. Two flavonoids from the Citrus genus, hesperidin (Hsd) and hesperetin (Hst), exhibit a range of biological activities, including antioxidant, anti-inflammatory, and anti-cancer effects³⁴. Also, it's possible that some strains of Gram+ and Gram-bacteria, including *Staphylococcus aureus*, are immune to the antimicrobial effects of glycoside flavones^{34,35}. The many effects of flavonoids, such as hesperidin, on healthy dentin have opened up a new avenue for the development of therapeutic medicines to support bonding stability on caries-affected dentin.

When Islam et al. (2012) looked at how adding natural cross-linkers affected the resin-dentine bond strength of a self-etching adhesive, they discovered that adding HPN significantly increased micro-TBS, adding grape seed extract (GSE) significantly decreased micro-TBS, and adding chlorhexidine (CHX) had no statistically significant differences³⁶. Their findings demonstrate that the micro-TBS and mechanical characteristics of the bonded interface were immediately enhanced by the addition of HPN to the Clearfil SE primer. Ghorab et al. (2018)

examined the antimicrobial activity and adhesive properties of a condensed total-etch adhesive system containing varying amounts of Hesperidin (HPN), coming to the conclusion that doing so significantly increased the dental adhesive's immediate TBS at 0.2 wt percent and 0.5 wt percent HPN (P 0.05)³². Thermocycling significantly reduced the TBS of dental adhesives with 0.5 wt percent HPN incorporated, which showed a potential antibacterial impact without changing the adhesive properties.

When combined with DMSO and added to a total etch bonding agent, hesperidin is very efficient against *L. acidophilus*, *S. mutans*, and *E. faecalis*³⁷. The adhesive characteristics of total etch dentin adhesive are not adversely affected when HPN is applied at 2% concentration, but a promising antibacterial effect is attained. Hesperidin contains collagen cross-linking and MMP inhibitory activities. As a result, dentin adhesive's initial bond strength may be improved^{38,39}. Hesperidin can be more easily dissolved with the use of dimethyl sulfoxide (DMSO). DMSO enhances adhesive penetration into the exposed collagen matrix after acid conditioning, which enhances adherence⁴⁰. In addition, DMSO has the power to inhibit the collagen-degrading enzymes that are obtained from the host. The hybrid layer's durability is enhanced as a result. Biocompatible adhesive systems are those that establish a solid bond with biological tissues and permit both tissue repair and tissue differentiation. In the current investigation, the cytotoxicity of the experimental adhesive was affected by its concentration. But those standards are still respectable. Hesperidin (HPN) functions as a natural collagen cross-linker that aids in guarding against exposed collagen deterioration brought on by acid etching. The degree of conversion of unreacted monomers is improved by its antioxidant property⁴¹. This naturally occurring flavonoid called Hesperidin has a chroman ring that can interact with proteins high in proline, such collagen, to cross-link them and enhance the mechanical qualities of dentin. As a result, it aids in enhancing the biocompatibility and sealing capacity of dentin adhesives.

CONCLUSION

Within the constraints, it can be said that adding 2% hesperidin to total etch dentin adhesive could achieve therapeutic objectives that are important for biocompatibility without impairing dentin adhesive's adhesive capabilities. Moreover, it is antibacterial. Hence, it can be utilised to prevent the development of secondary caries in adhesive restorations.

REFERENCES

1. Asl HRM, Asl EM. A review on restorative and preventive materials used in dentistry. *Journal of Dental Health, Oral Disorders & Therapy* 2018; 9: 526–529.
2. Zou Y, Jessop JLP, Armstrong SR. Apparent conversion of adhesive resin in the hybrid layer, Part II: In situ studies of the resin-dentin bond. *Journal of Biomedical Materials Research Part A* 2009; 89A: 355–362.
3. Walter R, Miguez PA, Arnold RR, et al. Effects of Natural Cross-Linkers on the Stability of Dentin Collagen and the Inhibition of Root Caries in vitro. *Caries Research* 2008; 42: 263–268.
4. Lührs A-K, Guhr S, Günay H, et al. Shear bond strength of self-adhesive resins compared to resin cements with etch and rinse adhesives to enamel and dentin in vitro. *Clinical Oral Investigations* 2010; 14: 193–199.
5. Jacobsen T, Söderholm K-JM, Garcea I, et al. Calcium leaching from dentin and shear bond strength after etching with phosphoric acid of different concentrations. *European Journal of Oral Sciences* 2000; 108: 247–254.
6. Vidal CMP, Tjäderhane L, Scaffa PM, et al. Abundance of MMPs and Cysteine Cathepsins in Caries-affected Dentin. *Journal of Dental Research* 2014; 93: 269–274.
7. Van Doren SR. Matrix metalloproteinase interactions with collagen and elastin. *Matrix Biology* 2015; 44-46: 224–231.
8. Abdullah H, Khodair Z, AL-Zanganawee J, et al. Study of inter laminate layer effect on Impact and Hardness Properties for Unsaturated Polyester resin reinforced Hybrid Fabric composite. *Journal of Garmian University* 2017; 4: 401–408.
9. Rudawska A. Adhesives: Applications and Properties. BoD – Books on Demand, 2016.
10. Wdowiak K, Walkowiak J, Pietrzak R, et al. Bioavailability of Hesperidin and Its Aglycone Hesperetin—Compounds Found in Citrus Fruits as a Parameter Conditioning the Pro-Health Potential (Neuroprotective and Antidiabetic Activity)—Mini-Review. *Nutrients* 2022; 14: 2647.
11. Adapa S, Sushanth VH, Prashant GM, et al. In vitro antimicrobial activity of *Spinacia Oleracea* against *Streptococcus mutans* and *Lactobacillus acidophilus*. *Journal of Indian Association of Public Health Dentistry* 2018; 16: 251.
12. Berto LA. Antimicrobial activity of plants from Brazilian Cerrado against *Streptococcus mutans*. DOI: 10.47749/t/unicamp.2014.927794.
13. Fujimori K, Arita A, Kumagai T. Effect of mechanical properties of adhesives on bond strength. *Dental Materials* 2019; 35: e15.
14. de Macedo FAA, Souza NO, Lemos MVS, et al. Dentin bonding and physicochemical properties of adhesives incorporated with epigallocatechin-3-gallate. *Odontology* 2019; 107: 23–28.
15. Malli Sureshbabu N, Selvarasu K, V JK, et al. Concentrated Growth Factors as an Ingenious Biomaterial in Regeneration of Bony Defects after Periapical Surgery: A Report of Two Cases. *Case Rep Dent* 2019; 2019: 7046203.
16. Ahad M, Gheena S. Awareness, attitude and knowledge about evidence based dentistry among the dental practitioner in Chennai city. *J Adv Pharm Technol Res* 2016; 9: 1863.
17. PradeepKumar AR, Shemesh H, Jothilatha S, et al. Diagnosis of Vertical Root Fractures in Restored Endodontically Treated Teeth: A Time-dependent Retrospective Cohort Study. *J Endod* 2016; 42: 1175–1180.
18. Jangid K, Alexander AJ, Jayakumar ND, et al. Ankyloglossia with cleft lip: A rare case report. *J Indian Soc Periodontol* 2015; 19: 690–693.
19. Kumar A, Sherlin HJ, Ramani P, et al. Expression of CD 68, CD 45 and human leukocyte antigen-DR in central and peripheral giant cell granuloma, giant cell tumor of long bones, and tuberculous granuloma: An immunohistochemical study. *Indian J Dent Res* 2015; 26: 295–303.
20. Manohar J, Abilasha R. A Study on the Knowledge of Causes and Prevalance of Pigmentation of Gingiva among Dental Students. *Indian Journal of Public Health Research & Development* 2019; 10: 95.
21. Sekar D, Mani P, Biruntha M, et al. Dissecting the functional role of microRNA 21 in osteosarcoma. *Cancer Gene Ther* 2019; 26: 179–182.
22. Girija SA, Jayaseelan VP, Arumugam P. Prevalence of VIM- and GIM-producing *Acinetobacter baumannii* from patients with severe urinary tract infection. *Acta Microbiol Immunol Hung* 2018; 65: 539–550.
23. Maheswari TNU, Venugopal A, Sureshbabu NM, et al. Salivary micro RNA as a potential biomarker in oral potentially malignant disorders:

- A systematic review. *Ci Ji Yi Xue Za Zhi* 2018; 30: 55–60.
24. Subashri A, Maheshwari TNU. Knowledge and attitude of oral hygiene practice among dental students. *J Adv Pharm Technol Res* 2016; 9: 1840.
 25. Sridharan G, Ramani P, Patankar S, et al. Evaluation of salivary metabolomics in oral leukoplakia and oral squamous cell carcinoma. *J Oral Pathol Med* 2019; 48: 299–306.
 26. Ezhilarasan D, Apoorva VS, Ashok Vardhan N. Syzygium cumini extract induced reactive oxygen species-mediated apoptosis in human oral squamous carcinoma cells. *J Oral Pathol Med* 2019; 48: 115–121.
 27. Mathew MG, Samuel SR, Soni AJ, et al. Evaluation of adhesion of Streptococcus mutans, plaque accumulation on zirconia and stainless steel crowns, and surrounding gingival inflammation in primary molars: randomized controlled trial. *Clin Oral Investig* 2020; 24: 3275–3280.
 28. Vijayashree Priyadharsini J. In silico validation of the non-antibiotic drugs acetaminophen and ibuprofen as antibacterial agents against red complex pathogens. *J Periodontol* 2019; 90: 1441–1448.
 29. Chandrasekar R, Chandrasekhar S, Sundari KKS, et al. Development and validation of a formula for objective assessment of cervical vertebral bone age. *Prog Orthod* 2020; 21: 38.
 30. Hass V, Liu H, Cook W, et al. Distinct effects of polyphenols and solvents on dentin collagen crosslinking interactions and biostability. *Dental Materials* 2021; 37: 1794–1805.
 31. Wu Y, Chai B, Wang L, et al. Antibacterial activity of total flavonoids from *Ilex rotunda* Thunb. and different antibacterials on different multidrug-resistant bacteria alone or in combination. DOI: 10.1101/457911.
 32. Ghorab S, Ibraheim A. Effect of hesperidin on antibacterial activity and adhesive properties of an etch-and-rinse adhesive system. *Egyptian Dental Journal* 2018; 64: 3801–3812.
 33. Tronchet JMJ, Zerelli S, Dolatshahi N, et al. Influence of the structure of the sugar moiety on the cytotoxic and antiviral properties of sugar electrophiles. *Chemical and Pharmaceutical Bulletin* 1988; 36: 3722–3725.
 34. Parhiz H, Roohbakhsh A, Soltani F, et al. Antioxidant and Anti-Inflammatory Properties of the Citrus Flavonoids Hesperidin and Hesperetin: An Updated Review of their Molecular Mechanisms and Experimental Models. *Phytotherapy Research* 2015; 29: 323–331.
 35. Johnson A. Activity of daptomycin against multi-resistant Gram-positive bacteria including enterococci and Staphylococcus aureus resistant to linezolid. *International Journal of Antimicrobial Agents* 2004; 24: 315–319.
 36. Islam S, Hiraishi N, Nassar M, et al. Effect of natural cross-linkers incorporation in a self-etching primer on dentine bond strength. *J Dent* 2012; 40: 1052–1059.
 37. Stape THS, Tjäderhane L, Szesz A, et al. DMSO improves long-term dentin bonding of etch-and-rinse and self-etch adhesives. *Dental Materials* 2015; 31: e27–e28.
 38. Camim F da S, da Silva Camim F. Influence of MMP inhibitors on bond strength of adhesive restorations: systematic review and meta-analysis. DOI: 10.11606/d.25.2019.tde-25112019-205524.
 39. Coli P, Alaeddin S, Wennerberg A, et al. In vitro dentin pretreatment, Surface roughness and adhesive shear bond strength. *European Journal of Oral Sciences* 1999; 107: 400–413.
 40. Zabeu GS. Use of dimethyl sulfoxide (DMSO) to optimize adhesive interface: analysis of its effectiveness on the mechanical and biological properties in dentin bonding. DOI: 10.11606/t.25.2021.tde-07022022-113825.
 41. Cao R, Zhao Y, Zhou Z, et al. Enhancement of the water solubility and antioxidant activity of hesperidin by chitoooligosaccharide. *Journal of the Science of Food and Agriculture* 2018; 98: 2422–2427.